

Identification of Intermediate in Evolutionary Model of Enterohemorrhagic *Escherichia coli* O157

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Highly pathogenic enterohemorrhagic *Escherichia coli* (EHEC) O157 cause a spectrum of clinical signs that include diarrhea, bloody diarrhea, and hemolytic uremic syndrome. The current evolutionary model of EHEC O157:H7/H⁻ consists of a stepwise evolution scenario proceeding from O55:H7 to a node (hypothetical intermediate) that then branches into sorbitol-fermenting (SF) O157:H⁻ and non-SF (NSF) O157:H7. To identify this hypothetical intermediate, we performed single nucleotide polymorphism analysis by sequencing of 92 randomly distributed backbone genomic regions of 40 O157:H7/H⁻ isolates. Overall, 111 single nucleotide polymorphisms were identified in 75/92 partial open reading frames after sequencing 51,041 nt/strain. The EHEC O157:H7 strain LSU-61 from deer occupied an intermediate position between O55:H7 and both O157 branches (SF and NSF O157), complementing the stepwise evolutionary model of EHEC O157:H7/H⁻. The animal origin of this intermediate emphasizes the value of nonhuman reservoirs in the clarification of the evolution of human pathogens.

Enterohemorrhagic *Escherichia coli* (EHEC) belongs to the Shiga toxin-producing *E. coli* group and causes clinical signs ranging from watery to bloody diarrhea for most symptomatically infected patients (1,2). EHEC serotypes O157:H7 and O157:H⁻ (nonmotile)

the most frequently isolated from patients with severe EHEC-associated diseases, such as bloody diarrhea and hemolytic uremic syndrome. Infections caused by EHEC O157:H7/H⁻ are major public health threats and require considerable resources for control and prevention (1,3). Sorbitol-fermenting (SF) EHEC O157:H⁻, initially found in Germany and later in other countries such as Scotland, Finland, and Australia, are increasingly associated with severe disease (4). These strains can ferment sorbitol after overnight incubation on sorbitol MacConkey agar, unlike non-SF (NSF) EHEC O157:H7. Today, SF EHEC O157:H⁻ strains cause ≈20% of all hemolytic uremic syndrome cases in Germany (4–8). Classic NSF EHEC O157:H7 are of animal origin and have caused multiple outbreaks through contaminated food (4), but SF EHEC O157:H⁻ are almost exclusively isolated from humans, which suggests that humans are the main reservoir (5).

On the basis of multilocus enzyme electrophoresis and multilocus sequence typing (MLST) data (9,10), the evolutionary model of EHEC O157 suggests that EHEC O157 emerged from *E. coli* O55:H7 by loss and acquisition of virulence and phenotypic traits (10). To further explain the evolution from O55:H7, a hypothetical intermediate and putatively extinct clone (missing link) SF O157:H7 emerging from O55:H7 was introduced; theoretically, it is from this intermediate that the 2 branches (NSF O157:H7 and SF O157:H⁻) diverged (9,10).

Shaikh and Tarr subdivided NSF O157:H7 into 3 clusters (11); in their analysis, the SF O157:H⁻ branch remained evolutionary conserved and clearly separated from NSF O157:H7, with additional data suggesting a hypothetical intermediate. Recent studies based on whole

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core genome single-nucleotide polymorphisms (SNPs) enabled precise reconstruction of this model (12). The *E. coli* O157:H⁻ strain LSU-61, which was isolated from a deer (10,13), had been previously discussed by Feng et al. as a potential intermediate, but that hypothesis was rejected because the strain lacked a gene encoding Shiga toxin (*stx*) and had a distinct MLST sequence type (10). We used an SNP-based approach to examine isolates from different sources of EHEC O157:H7/H⁻ to further elucidate the evolutionary model of emergence of this pathogen, paying particular attention to identifying the “missing link” hypothetical intermediate.

Materials and Methods

Bacterial Strains Analyzed

Of the 50 EHEC strains examined (Table), 48 were serotype O157:H7/H⁻ and 2 were O55:H7. Core or complete genome sequences were available for 8 O157 and 2 O55:H7 strains; these sequences served as a framework of the evolutionary model of EHEC O157. The remaining 40 strains consisted of 13 O157:H7/H⁻ strains that represented different clusters according to previous multilocus variable-number tandem-repeat analysis (19); 26 O157:H7/H⁻ strains isolated during 1987–2010 that were randomly chosen from our strain collection; and strain LSU-61, which was considered to be an intermediate (10).

Identification of EHEC O157 Strains

All 39 EHEC O157 isolates from our laboratory were isolated from stool samples as described (20,21). Isolates were confirmed to be *E. coli* by the API 20 Etest (bioMérieux, Marcy l’Etoile, France) and serotyped by using antiserum against *E. coli* O antigens 1–181 and H antigens 1–56 (22). Subtyping of *fliC* genes in nonmotile isolates by using *Hha*I restriction fragment-length polymorphism of amplicons obtained with primers FSa1 and rFSa1 (23,24) confirmed the presence of *fliC*_{H7} in all isolates. All strains were frozen at –70°C until further use.

Isolation of DNA

A single colony from a fresh overnight culture on Columbia blood agar (Heipha, Eppelheim, Germany) was inoculated into a liquid culture of nutrient broth medium (Heipha) and incubated overnight at 37°C. The liquid culture was used to prepare DNA as described (25), except that phenol extraction was omitted and the corresponding supernatants were directly precipitated with isopropanol.

Cluster Classification of O157:H7 Strains

Previously determined SNP patterns T/G/T/A or G/T/C/C at Sakai genome positions 337,933 (ECs0320, putative receptor), 1,460,599 (ECs1414, curli production assembly

transport component), 2,370,797 (ECs2397, transport system permease protein), and 5,404,166 (ECs5279, *fimH*-locus) have been shown to be cluster specific (12). On this basis, we used Sanger sequencing to group strains into cluster 3 or cluster 1 of subgroup C. Because the prototype strain of cluster 2 shared the SNP pattern with cluster 3, strains of cluster 2 were differentiated by using the published cluster differentiation scheme based on the occupancy of *stx* integration sites and the *stx* genotype (11,15). SNP pattern T/G/T/C was declared as unknown.

MLST and Sequencing of EHEC O157

Core Genomic Loci

As a first classification, we used MLST to determine the sequence type (ST) for all prototype strains of each subgroup and cluster by sequencing internal fragments of 7 housekeeping genes (*adk*, *fumC*, *gyrB*, *icd*, *mdh*, *purA*, and *recA*) (26). Alleles, STs, and clonal complexes were assigned in accordance with the *E. coli* MLST website (<http://mlst.ucc.ie/mlst/dbs/Ecoli>).

In addition to 10 SNP localizations that were known to differentiate subgroups and clusters (12), we randomly selected 82 additional backbone genomic regions for more in-depth SNP analysis. Using the Primer3 algorithm (<http://frodo.wi.mit.edu/primer3>), we developed 93 primer pairs that generated PCR products of backbone genomic regions ranging from 600 to 700 bp (online Technical Appendix Table 1, wwwnc.cdc.gov/EID/pdfs/11-1414-Techapp.pdf); for open reading frame (ORF) ECs3076, 2 separate primer pairs were designed to cover 2 described SNP localizations (12). EHEC O157:H7 strain Sakai served as a reference (GenBank accession no. NC_002655).

PCR was performed in a 14-μL reaction mixture containing 7 μL REDTaq (Sigma Aldrich, St. Louis, MO, USA), ≈6 ng DNA, and 1.5 μL each forward and reverse primer, with a final concentration of 10 μmol/L. The cycling reaction conditions were initial denaturation (2 min at 94°C), 35 cycles of denaturation (45 s at 94°C), annealing (60 s at 60°C), and extension (90 s at 72°C), followed by a final extension (10 min at 72°C). PCR products were purified by using the exonuclease I (New England Biolabs GmbH, Frankfurt-Höchst, Germany) and shrimp alkaline phosphatase (USB Amersham, Freiburg, Germany) according to methods modified from (27). In brief, 7 μL of the PCR product was incubated simultaneously with 1.5 U of each enzyme at 37°C for 45 min, followed by enzyme heat inactivation at 80°C.

For sequencing of both strands, 2 μL of the purified amplicons was mixed with 0.5 μL premix from the ABI Prism BigDye Terminator v3.1 Ready Reaction Cycle Sequencing Kit (Applied Biosystems, Foster City, CA, USA) plus 1.8 μL Tris-HCl-MgCl₂ buffer (400 mmol/L Tris-HCl, 10 mmol/L MgCl₂; pH 9) and 2 μL (10 μmol/L)

from the sequencing primer (forward or reverse primer, in a total volume of 10 µL. The cycling reaction conditions were 25 cycles of denaturation (10 s at 96°C) and combined annealing and extension (4 min at 60°C). Finally, the sequencing reaction products were purified by using an alcohol precipitation method as recommended by the

manufacturer and loaded onto a 3130xl Genetic Analyzer (Applied Biosystems) for capillary sequencing.

Genotypic Characterization of LSU-61

To further evaluate the genotype of LSU-61 and its potential role in the evolutionary model of EHEC O157,

Table. Fifty strains used for SNP typing of enterohemorrhagic *Escherichia coli* O157*

Strain ID	Year of isolation	Illness	SF status and serotype	Subgroup/cluster†	Reference and/or GenBank accession no.
TB182A‡	1991	D	SF O55:H7	A	(12)
CB9615‡	2003	D	SF O55:H7	A	NC_013941
493/89‡	1989	HUS	SF O157:H ⁻	B	(16)
87-14‡	1987	HUS	NSF O157:H7	C1	(12)
EC4115‡	2006	BD	NSF O157:H7	C1	NC_011353
TW14359‡	2006	BD	NSF O157:H7	C1	NC_013008 (12,17)
TW14588‡	2006	BD	NSF O157:H7	C3	NZ_ABKY00000000.2
86-24‡	1986	HUS	NSF O157:H7	C2	(12)
Sakai‡	1996	D	NSF O157:H7	C3	NC_002695 (18)
EDL933‡	1983	NA	NSF O157:H7	C3	(14)
LSU-61	2001	NA	SF O157:H7	Unknown intermediate	(10,13)
SNPO157_01	1987	HUS	NSF O157:H7	C1	This study
SNPO157_02	1988	D	NSF O157:H7	C1	This study
SNPO157_03	1988	HUS	SF O157:H ⁻	B	This study
SNPO157_04	1990	HUS	NSF O157:H7	C1	This study
SNPO157_05	1991	HUS	NSF O157:H ⁻	C3	This study
SNPO157_06	1992	HUS	NSF O157:H7	C3	This study
SNPO157_07	1993	HUS	NSF O157:H7	C1	This study
SNPO157_08	1995	HUS	NSF O157:H7	C1	This study
SNPO157_09	1995	HUS	SF O157:H ⁻	B	This study
SNPO157_10	1996	HUS	NSF O157:H ⁻	C1	This study
SNPO157_11	1996	HUS	SF O157:H ⁻	B	This study
SNPO157_12	1996	HUS	SF O157:H ⁻	B	This study
SNPO157_13	1996	HUS	NSF O157:H7	C1	This study
SNPO157_14	1997	HUS	NSF O157:H7	C1	This study
SNPO157_15	1997	HUS	NSF O157:H7	C1	This study
SNPO157_16	1998	HUS	NSF O157:H7	C1	This study
SNPO157_17	1999	HUS	NSF O157:H7	C1	This study
SNPO157_18	1999	HUS	NSF O157:H7	C3	This study
SNPO157_19	2000	HUS	NSF O157:H7	C1	This study
SNPO157_20	2000	D	NSF O157:H7	C3	This study
SNPO157_21	2001	HUS	SF O157:H ⁻	B	This study
SNPO157_22	2001	HUS	NSF O157:H ⁻	C3	This study
SNPO157_23	2002	D	NSF O157:H7	C3	This study
SNPO157_24	2002	A	NSF O157:H7	C1	This study
SNPO157_25	2003	HUS	NSF O157:H7	C3	This study
SNPO157_26	2004	HUS	NSF O157:H7	C1	This study
SNPO157_27	2005	HUS	NSF O157:H7	C3	This study
SNPO157_28	2005	BD	NSF O157:H7	C3	This study
SNPO157_29	2005	HUS	NSF O157:H7	C1	This study
SNPO157_30	2006	HUS	NSF O157:H7	C3	This study
SNPO157_31	2007	HUS	NSF O157:H7	C3	This study
SNPO157_32	2007	HUS	NSF O157:H7	C3	This study
SNPO157_33	2008	HUS	NSF O157:H7	C3	This study
SNPO157_34	2008	D	SF O157:H ⁻	B	This study
SNPO157_35	2008	HUS	SF O157:H ⁻	B	This study
SNPO157_36	2009	D	NSF O157:H7	§	This study
SNPO157_37	2009	HUS	SF O157:H ⁻	B	This study
SNPO157_38	2010	HUS	NSF O157:H7	C3	This study
SNPO157_39	2010	HUS	SF O157:H ⁻	B	This study

*All strains were isolated from humans except strain LSU-61, which was isolated from a deer (10), and EDL933, which was isolated from food (14).

Strains isolated from humans were categorized into 3 subgroups (11,12); subgroup A represents isolates of serotype O55:H7, subgroup B SF O157:H- isolates, and subgroup C NSF O157:H7. Subgroup C is subdivided into clusters 1–3. SNP, single-nucleotide polymorphism; ID, identification; SF, sorbitol fermenting; D, diarrhea; HUS, hemolytic uremic syndrome; NSF, non-SF; BD, bloody diarrhea; NA, not applicable; A, asymptomatic.

†Subgroup and, if applicable, cluster designation based on 4 SNP loci (Sakai genome positions 337933, 1460599, 2370797, and 5404166) and the occupancy of potential *stx* integration sites in accordance with (11,12,15). **Boldface** indicates cluster designation of prototype strains.

‡Strains were analyzed in silico.

§SNP pattern for NSF O157:H7 grouping resulted in an unknown combination.

we investigated known *stx*-phage integration sites. We used the draft genome sequence of the O157 strain LSU-61 (GenBank accession no. AEUC00000000) (28). *yehV*, a known integration site of *stx1*, was screened in silico by using primer pair A/B from (29). For analysis of the *wrbA* locus, a site of integration of the *stx2* bacteriophage, we used primer pair C/D from (29). The 2 other currently known potential integration sites of *stx2*, *yecE* and *sbcB*, were screened by using primer pairs EC10/EC11, *yecD*-fwd/*yecN*-rev, and *sbcB1*/*sbcB2* (30).

Data Analysis

Sequence trace files were analyzed and stored by using SeqSphere software version 0.9 beta (Ridom GmbH, Münster, Germany); a minimum-spanning tree was constructed with the integrated minimum-spanning tree algorithm. Gene functions were categorized by using the Pathosystems Resource Integration Center database (www.patricbrc.org/portal/portal/patric/Home) and corresponding Kyoto Encyclopedia of Genes and Genomes (KEGG; www.genome.jp/kegg) assignments. Overall, genes were grouped into 3 functional categories: metabolism/housekeeping, putative metabolism/housekeeping, and hypothetical protein. If no KEGG phenotype assignment was found, a putative metabolism/housekeeping function was predicted on the basis of BLAST (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) results.

Results

Of the 48 EHEC O157 strains studied, 10 were SF serotype O157:H⁻. Subgrouping and cluster designation of the NSF O157 strains resulted in 18 cluster 1 strains, 1 cluster 2 strain, and 17 cluster 3 strains. For 2 strains, LSU-61 and SNPO157_36, no characteristic SNP pattern was determined (Table). Further characterization by MLST of prototype strains that defined subgroups and clusters resulted in identical STs for all SF and NSF O157 (ST11) and in closely related STs of the O55:H7 strains (ST335).

The 50 strains of serotypes O157:H7/H⁻ and O55:H7 were further characterized with respect to their SNP prevalence in the core genome. In total, 92 core genomic loci were analyzed, comprising 51,041 bp sequencing information ($\approx 0.9\%$ of the O157:H7 Sakai genome) (Table; online Technical Appendix Table 2). Sequencing demonstrated 111 biallelic variants, an average of 1.2 variants per sequenced locus (online Technical Appendix Table 3). Deletions or insertions were not detected.

Of the 111 SNPs, 53 (47.7%) were synonymous SNPs (sSNPs) and 58 (52.3%) were nonsynonymous SNPs (nsSNPs); 78 (70.3%) SNPs were transitions, and 33 (29.7%) were transversions. Concatenation of all loci resulted in an average of 1 SNP every 460 bp; sSNPs occurred every 963 bp, and nsSNPs every 880 bp. On

the level of analyzed partial ORFs, 1 SNP was found in 45 partial ORFs, 2 SNPs in 25 partial ORFs, 3 SNPs in 4 partial ORFs, and 4 SNPs in 1 partial ORF.

To further elucidate the SNP distribution, we categorized the 92 loci into 3 functional groups. Most loci belonged to (putative) metabolism or housekeeping genes because these were chosen solely from backbone regions. If no KEGG assignment was possible, we estimated the function of the corresponding fragment on the basis of BLAST homologies. Defined annotation information regarding the function in metabolism or housekeeping was determined for 25 partial ORFs. A housekeeping/metabolism function was predicted for 58 loci. The remaining 9 loci were hypothetical proteins only (online Technical Appendix).

On the basis of the 111 SNPs, the 50 strains were clustered into 27 SNP genotypes (Figure). The 111 SNPs were able to reconstruct the well-known evolutionary model (9–12) with the stepwise evolution from O55:H7 (subgroup A) to either SF O157:H⁻ (subgroup B) or NSF O157:H7/H⁻ (subgroup C, clusters 1–3). Strain LSU-61 interlinked all 3 subgroups, thereby substituting for the unknown intermediate (Figure). Moreover, the applied SNP scheme differentiated the O55:H7 strains and exhibited 5 genotypes within the 10 SF O157:H⁻ strains segregated from the SF O157:H7 strain LSU-61. The NSF O157:H7 subgroup C₁ strains comprising 18 isolates were differentiated in 9 SNP genotypes, and the subgroup C₃ strains (n = 17) exhibited 8 SNP genotypes.

To further validate the role of *stx*-negative LSU-61 as a potential intermediate, we investigated each known potential *stx* insertion site in silico to determine the presence or absence of a Shiga toxin–carrying bacteriophage. We conducted BLAST searches within the recently published draft genome sequence of LSU-61 (28) by using published primers for the different insertion sites (29,30). All insertion sites for *stx1* (*yehV*) and *stx2* (*wrbA*, *yecE*, *sbcB*) were intact.

To investigate the effect of selective pressure on some loci and potential selecting biases, we analyzed sSNP and nsSNP types separately. In each scenario, the phylogenetic reconstruction resulted in comparable branching, with distinct lineages for SF and NSF O157 and strain LSU-61 as an intermediate. Only the number of SNP genotypes differed slightly: 19 sSNP genotypes (13 NSF O157:H7, 3 SF O157:H⁻, 2 O55:H7, and LSU-61) based on the 53 sSNPs and 22 nsSNP genotypes (16 NSF O157:H7, 3 SF O157:H⁻, 2 O55:H7, and LSU-61) based on the 58 nsSNPs. This excludes strong selection bias of the different loci.

Discussion

On the basis of SNP analysis of 92 chromosomal backbone regions of EHEC O157, we identified an SF

O157:H7 strain that complements the current model of the stepwise evolution from O55:H7 to EHEC O157 in which the hypothetical intermediate between O55:H7 and SF and NSF O157:H7/H⁻ has been unknown (10,12). As with the highly human pathogenic O157:H7 lineage of EHEC, which is known to reside in cattle, deer, and other ruminants, this intermediate strain was isolated from a deer (13). These findings support previous observations (31,32) and suggest an evolution toward an animal reservoir for O157:H7 soon after O157:H⁻ and O157:H7 divergence. Strain LSU-61 is motile (H-phase 7) and enterohemolysin active (10), traits that are typical for NSF O157, further suggesting the intermediate character of LSU-61 between SF and NSF O157. In contrast to the MLST scheme applied from Feng et al. (10) and Lacher et al. (33), our MLST analysis, using the scheme of Wirth et al. (26) that analyzes different genes, further corroborates the intermediate character of LSU-61 because it shares the same ST with SF (subgroup B prototype strain) and NSF O157 (subgroup C prototype strains).

Strain LSU-61 does not carry a *stx* gene, but this fact does not contradict our findings because these genes are encoded on bacteriophages that can be acquired and lost (30,34,35), and we do not have evidence of a progenitor to LSU-61 that contains *stx* genes. Although known potential *stx* phage integration sites in O157 were intact, the possibility of a previous *stx* bacteriophage carriage cannot be excluded. If the SF O157:H7 cluster emerged ≈3,000–4,000 years ago (12), certain genetic and phenotypic changes (10) occurred well before the first descendants of this cluster were isolated and characterized.

Two previous studies (31,32) reported isolated comparable strains to LSU-61 from (European) red deer, belonging to the same family (*Cervidae*) as white-tailed deer (North America), with comparable phenotypic and genotypic traits. Some of these were SF O157:H7 strains (*stx* negative or positive, β-glucuronidase positive activity) (31,32). The proof of the existence of SF O157:H7 in a ruminant (deer) host may indicate transfer into animals soon after the 2 (human pathogenic) O157 subgroups B and C emerged. On the basis of shared characteristics with both O157 branches, we suggest strain LSU-61 as a representative of the intermediate cluster complementing the stepwise evolutionary model of EHEC O157. The phylogeny based on either sSNPs or nsSNPs also resulted in a comparable phylogenetic tree with LSU-61 as a member of the progenitor node, underlining its intermediate role.

On the level of gene categories, a higher percentage of sSNPs, though fewer SNPs overall, were observed in the metabolism/housekeeping category compared with the putative metabolism/housekeeping category. The higher rate of nsSNPs in the latter category, resulting in a higher

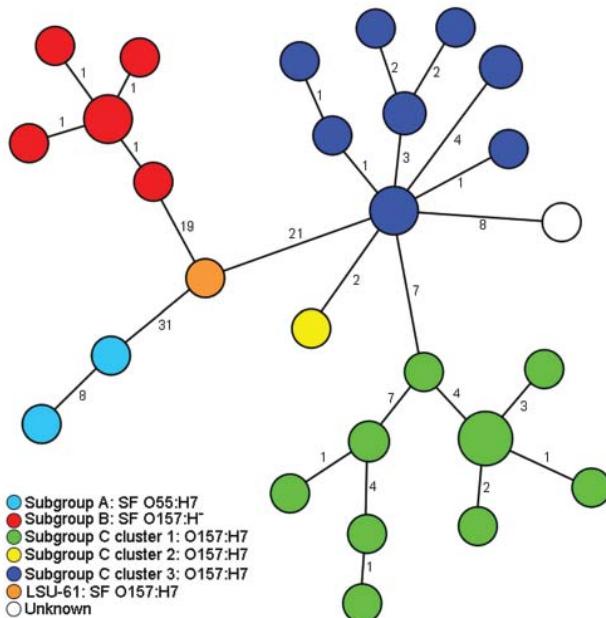


Figure. Minimum-spanning tree based on single-nucleotide polymorphism (SNP) genotypes illustrating the phylogeny of 50 enterohemorrhagic *Escherichia coli* O157:H7/H⁻ and O55:H7 isolates and the intermediate position of strain LSU-61 during the evolution of O157. Each node represents a unique SNP genotype. The size of each node is proportional to the number of isolates per SNP genotype based on sequence analysis of 51,041 bp comprising 92 partial open reading frames. Numbers on lines between nodes represent distances between the nodes, i.e., the number of SNPs. The node size is proportional to the number of strains sharing the same genotype. Strains are colored according to their classification into subgroups and clusters based on information from Saikh and Tarr (11) and Leopold et al. (12). Strain LSU-61 represents a potential intermediate interlinking all 3 subgroups. SF, sorbitol fermenting.

phenotypic diversity, might be explained by uncertain gene categorization because of currently limited knowledge of gene function. Therefore, SNP typing results may help to find genes involved in host-pathogen interactions rather than in metabolism or housekeeping only. SNP data for hypothetical proteins are difficult to interpret because information about their function is too imprecise to enable estimation of the effect of evolutionary pressure.

The fact that 35 of the 38 O157:H7 strains were subgrouped into either cluster 1 or 3 (17 and 18 strains, respectively) shows a certain persistence of these O157:H7 clusters (29), characterized by a successful pathogenicity, for example, outbreaks over a broad time frame (4). The preponderance of cluster 1 strains has been noted before, as have the paucity of cluster 2 and the diminished proportion of cluster 3 strains in North America (29). We observed a higher number of SNPs within the different NSF O157:H7 clusters compared with the few SNPs within restricted SF

O157:H⁻ genotypes and a maximum pairwise distance of 2 SNPs (Figure). A reason for this phenomenon may be the different animal host origins for the NSF O157:H7 clade, whereas SF O157:H⁻ are considered to have only 1 main host, humans (5,19). This high conservation was similarly recognized when multilocus variable-number tandem-repeat analysis was applied (19). In this context, certain SNP genotypes may serve to illuminate several strain-specific characteristics, such as increased virulence and other phenotypic traits, as other studies have similarly observed for both SF and NSF O157 (36,37).

Our results could be interpreted as if C₂ strain 86-24 is an offshoot of cluster 3, which is in contrast to the established stepwise model of O157. However, we believe that this is an artifact caused by sampling bias of the investigated 92 loci because only 11 backbone SNPs have been found to differentiate cluster 2 and 3 within the whole chromosomal backbone (12). One strain (SNPO157_36) did not cluster into any known O157:H7 cluster (Figure).

In summary, our identification of an intermediate member of the EHEC 1 clade complements the current evolutionary model of EHEC O157 by using chromosomal backbone SNP data of a spatiotemporally diverse strain collection. The different levels of genotypic conservation within the subgroups and the animal origin of the intermediate underline the great effect of host-pathogen interaction on the evolution of bacterial species. Future studies should focus on this interaction within both human and animal hosts to understand the evolution and persistence in nature of such human pathogens. The survival of the ancestral pathogen until today suggests that its genetic attributes could be informative in identifying fitness and potentially pathogenic loci.

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References

- Holtz LR, Neill MA, Tarr PI. Acute bloody diarrhea: a medical emergency for patients of all ages. *Gastroenterology*. 2009;136:1887-98. <http://dx.doi.org/10.1053/j.gastro.2009.02.059>
- Levine MM. *Escherichia coli* that cause diarrhea: enterotoxigenic, enteropathogenic, enteroinvasive, enterohemorrhagic, and enteroadherent. *J Infect Dis*. 1987;155:377-89. <http://dx.doi.org/10.1093/infdis/155.3.377>
- Tarr PI, Gordon CA, Chandler WL. Shiga toxin-producing *Escherichia coli* and haemolytic uraemic syndrome. *Lancet*. 2005;365:1073-86. [http://dx.doi.org/10.1016/S0140-6736\(05\)71144-2](http://dx.doi.org/10.1016/S0140-6736(05)71144-2)
- Karch H, Tarr PI, Bielaszewska M. Enterohaemorrhagic *Escherichia coli* in human medicine. *Int J Med Microbiol*. 2005;295:405-18. <http://dx.doi.org/10.1016/j.ijmm.2005.06.009>
- Karch H, Bielaszewska M. Sorbitol-fermenting Shiga toxin-producing *Escherichia coli* O157:H⁻ strains: epidemiology, phenotypic and molecular characteristics, and microbiological diagnosis. *J Clin Microbiol*. 2001;39:2043-9. <http://dx.doi.org/10.1128/JCM.39.6.2043-2049.2001>
- Karch H, Mellmann A, Bielaszewska M. Epidemiology and pathogenesis of enterohaemorrhagic *Escherichia coli*. *Berl Munch Tierarztl Wochenschr*. 2009;122:417-24.
- Pennington H. *Escherichia coli* O157. *Lancet*. 2010;376:1428-35. [http://dx.doi.org/10.1016/S0140-6736\(10\)60963-4](http://dx.doi.org/10.1016/S0140-6736(10)60963-4)
- Werber D, Bielaszewska M, Frank C, Stark K, Karch H. Watch out for the even eviler cousin—sorbitol-fermenting *E. coli* O157. *Lancet*. 2011;377:298-9. [http://dx.doi.org/10.1016/S0140-6736\(11\)60090-1](http://dx.doi.org/10.1016/S0140-6736(11)60090-1)
- Feng P, Lampel KA, Karch H, Whittam TS. Genotypic and phenotypic changes in the emergence of *Escherichia coli* O157:H7. *J Infect Dis*. 1998;177:1750-3. <http://dx.doi.org/10.1086/517438>
- Feng PCH, Monday SR, Lacher DW, Allison L, Siitonen A, Keys C, et al. Genetic diversity among clonal lineages within *Escherichia coli* O157:H7 stepwise evolutionary model. *Emerg Infect Dis*. 2007;13:1701-6.
- Shaikh N, Tarr PI. *Escherichia coli* O157:H7 Shiga toxin-encoding bacteriophages: integrations, excisions, truncations, and evolutionary implications. *J Bacteriol*. 2003;185:3596-605. <http://dx.doi.org/10.1128/JB.185.12.3596-3605.2003>
- Leopold SR, Magrini V, Holt NJ, Shaikh N, Mardis ER, Cagno J, et al. A precise reconstruction of the emergence and constrained radiations of *Escherichia coli* O157 portrayed by backbone concatenomic analysis. *Proc Natl Acad Sci U S A*. 2009;106:8713-8.
- Dunn JR, Keen JE, Moreland D, Alex T. Prevalence of *Escherichia coli* O157:H7 in white-tailed deer from Louisiana. *J Wildl Dis*. 2004;40:361-5.
- Perna NT, Plunkett G III, Burland V, Mau B, Glasner JD, Rose DJ, et al. Genome sequence of enterohaemorrhagic *Escherichia coli* O157:H7. *Nature*. 2001;409:529-33. <http://dx.doi.org/10.1038/35054089>
- Shaikh N, Holt NJ, Johnson JR, Tarr PI. Fim operon variation in the emergence of enterohemorrhagic *Escherichia coli*: an evolutionary and functional analysis. *FEMS Microbiol Lett*. 2007;273:58-63. <http://dx.doi.org/10.1111/j.1574-6968.2007.00781.x>
- Karch H, Wiss R, Gloning H, Emmrich P, Alekseev S, Bockemühl J. Hemolytic-uremic syndrome in infants due to verotoxin-producing *Escherichia coli* [in German]. *Dtsch Med Wochenschr*. 1990;115:489-95. <http://dx.doi.org/10.1055/s-2008-1065036>
- Kulasekara BR, Jacobs M, Zhou Y, Wu Z, Sims E, Saenphimmachak C, et al. Analysis of the genome of the *Escherichia coli* O157:H7 2006 spinach-associated outbreak isolate indicates candidate genes that may enhance virulence. *Infect Immun*. 2009;77:3713-21. <http://dx.doi.org/10.1128/IAI.00198-09>
- Hayashi T, Makino K, Ohnishi M, Kurokawa K, Ishii K, Yokoyama K, et al. Complete genome sequence of enterohemorrhagic *Escherichia coli* O157:H7 and genomic comparison with a laboratory strain K-12. *DNA Res*. 2001;8:11-22. <http://dx.doi.org/10.1093/dnares/8.1.11>

19. Jenke C, Harmsen D, Weniger T, Rothgänger J, Hyytiä-Trees E, Bielaszewska M, et al. Phylogenetic analysis of enterohemorrhagic *Escherichia coli* O157, Germany, 1987–2008. *Emerg Infect Dis*. 2010;16:610–6.
20. Friedrich AW, Bielaszewska M, Zhang W, Pulz M, Kuczius T, Ammon A, et al. *Escherichia coli* harboring Shiga toxin 2 gene variants: frequency and association with clinical symptoms. *J Infect Dis*. 2002;185:74–84. <http://dx.doi.org/10.1086/338115>
21. Mellmann A, Bielaszewska M, Zimmerhackl LB, Prager R, Harmsen D, Tschaep H, et al. Enterohemorrhagic *Escherichia coli* in human infection: in vivo evolution of a bacterial pathogen. *Clin Infect Dis*. 2005;41:785–92. <http://dx.doi.org/10.1086/432722>
22. Prager R, Strutz U, Fruth A, Tschaep H. Subtyping of pathogenic *Escherichia coli* strains using flagellar (H)-antigens: serotyping versus *fliC* polymorphisms. *Int J Med Microbiol*. 2003;292:477–86. <http://dx.doi.org/10.1078/1438-4221-00226>
23. Sonntag AK, Prager R, Bielaszewska M, Zhang W, Fruth A, Tschaep H, et al. Phenotypic and genotypic analyses of enterohemorrhagic *Escherichia coli* O145 strains from patients in Germany. *J Clin Microbiol*. 2004;42:954–62. <http://dx.doi.org/10.1128/JCM.42.3.954-962.2004>
24. Zhang Y, Laing C, Steele M, Ziebell K, Johnson R, Benson AK, et al. Genome evolution in major *Escherichia coli* O157:H7 lineages. *BMC Genomics*. 2007;8:121. <http://dx.doi.org/10.1186/1471-2164-8-121>
25. Wilson K. Preparation of genomic DNA from bacteria. *Curr Protoc Mol Biol*. 2001;Chapter 2:Unit 2.4.
26. Wirth T, Falush D, Lan R, Colles F, Mensa P, Wieler LH, et al. Sex and virulence in *Escherichia coli*: an evolutionary perspective. *Mol Microbiol*. 2006;60:1136–51. <http://dx.doi.org/10.1111/j.1365-2958.2006.05172.x>
27. Dugan KA, Lawrence HS, Hares DR, Fisher CL, Budowle B. An improved method for post-PCR purification for mtDNA sequence analysis. *J Forensic Sci*. 2002;47:811–8.
28. Rump LV, Strain EA, Cao G, Allard MW, Fischer M, Brown EW, et al. Draft genome sequences of six *Escherichia coli* isolates from the stepwise model emergence of *Escherichia coli* O157:H7. *J Bacteriol*. 2011;193:2058–9. <http://dx.doi.org/10.1128/JB.00118-11>
29. Besser TE, Shaikh N, Holt NJ, Tarr PI, Konkel ME, Malik-Kale P, et al. Greater diversity of Shiga toxin-encoding bacteriophage insertion sites among *Escherichia coli* O157:H7 isolates from cattle than in those from humans. *Appl Environ Microbiol*. 2007;73:671–9. <http://dx.doi.org/10.1128/AEM.01035-06>
30. Bielaszewska M, Prager R, Zhang W, Friedrich AW, Mellmann A, Tschaep H, et al. Chromosomal dynamism in progeny of outbreak-related sorbitol-fermenting enterohemorrhagic *Escherichia coli* O157:NM. *Appl Environ Microbiol*. 2006;72:1900–9. <http://dx.doi.org/10.1128/AEM.72.3.1900-1909.2006>
31. García-Sánchez A, Sanchez S, Rubio R, Pereira G, Alonso JM, Hermoso de Mendoza J, et al. Presence of Shiga toxin-producing *E. coli* O157:H7 in a survey of wild artiodactyls. *Vet Microbiol*. 2007;121:373–7. <http://dx.doi.org/10.1016/j.vetmic.2006.12.012>
32. Diaz S, Vidal D, Herrera-Leon S, Sanchez S. Sorbitol-fermenting, β -glucuronidase-positive, Shiga toxin-negative *Escherichia coli* O157:H7 in free-ranging red deer in south-central Spain. *Foodborne Pathog Dis*. 2011;8:1313–5. <http://dx.doi.org/10.1089/fpd.2011.0923>
33. Lacher DW, Steinsland H, Blank TE, Donnenberg MS, Whittam TS. Molecular evolution of typical enteropathogenic *Escherichia coli*: clonal analysis by multilocus sequence typing and virulence gene allelic profiling. *J Bacteriol*. 2007;189:342–50. <http://dx.doi.org/10.1128/JB.01472-06>
34. Bielaszewska M, Köck R, Friedrich AW, von Eiff C, Zimmerhackl LB, Karch H, et al. Shiga toxin-mediated hemolytic uremic syndrome: time to change the diagnostic paradigm? *PLoS ONE*. 2007;2:e1024. <http://dx.doi.org/10.1371/journal.pone.0001024>
35. Mellmann A, Lu S, Karch H, Xu J, Harmsen D, Schmidt MA, et al. Recycling of Shiga toxin 2 genes in sorbitol-fermenting enterohemorrhagic *Escherichia coli* O157:NM. *Appl Environ Microbiol*. 2008;74:67–72. <http://dx.doi.org/10.1128/AEM.01906-07>
36. Manning SD, Motiwala AS, Springman AC, Qi W, Lacher DW, Ouellette LM, et al. Variation in virulence among clades of *Escherichia coli* O157:H7 associated with disease outbreaks. *Proc Natl Acad Sci U S A*. 2008;105:4868–73. <http://dx.doi.org/10.1073/pnas.0710834105>
37. Zhang W, Qi W, Albert TJ, Motiwala AS, Alland D, Hyytiä-Trees EK, et al. Probing genomic diversity and evolution of *Escherichia coli* O157 by single nucleotide polymorphisms. *Genome Res*. 2006;16:757–67. <http://dx.doi.org/10.1101/gr.4759706>

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FIG 3. Clinical relevance of pulmonary nontuberculous mycobacterium (NTM) isolates, Asia, 1971–2007. Relevance per species was defined as percentage of patients with pulmonary NTM isolates meeting the American Thoracic Society criteria. Species reported infrequently, i.e., <5%, are not shown. Data from 16,17,21,22,25,29,32,33.

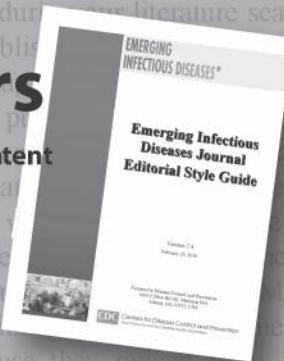
Style Guide for Authors

Revised. More information. Friendlier format. Searchable content **dy** supports their conclusion of the predominance of **AC** in **Asiats** their conclusion of the predominance of **other parts of the world, namely, North America and most parts of Europe (3).**

Third, we found that in some regions in Asia, RGM is a major cause of pulmonary NTM disease. This finding contrasts with our findings in Europe and North America. In a surveillance study from the Netherlands for instance, RGM caused only 3% of all pulmonary NTM

is the language restriction. The inclusion of languages other than English would probably have increased prevalence. For instance, during our literature search we came across 5 articles, published in Chinese, in PubMed, or Google Scholar. These aspects of the journal were already planned and developed. Nevertheless, our intention remains to include publications from all over the world, in all species in Asia and Europe. The span of the journal is limited by the language restriction.

Another limitation is the lack of studies from Asia. Because they ranged from 1969 to 2007, the studies did not cover the history, culture, and identification methods. Data should therefore be considered with caution because of the variety of laboratory procedures used.



Identification of Intermediate in Evolutionary Model of Enterohemorrhagic *Escherichia coli* O157

Technical Appendix

This appendix contains a detailed list of the 92 open reading frames (ORFs) (Technical Appendix Table 1) and single nucleotide polymorphisms (SNPs) (Technical Appendix Table 2) of all 50 strains of Enterohemorrhagic *Escherichia coli* O157 investigated in this study. The gene product, functional category, primer sequences, PCR product sizes, and number of base pairs analyzed are given for each ORF, and for each strain, SNP genotype is given at the 111 chromosomal SNP localizations determined in the study. Technical Appendix Table 3 lists all 111 SNPs discovered in this study, with detailed information about the SNP position within the ORF and its absolute position within the reference genome of enterohemorrhagic *E. coli* O157:H7 strain Sakai (GenBank accession no. NC_002695). The reference nucleotide, its variant, and the corresponding codons are listed in addition to the codon effect (synonymous or non-synonymous mutation).

Technical Appendix Table 1. Detailed list of the 92 ORFs investigated

ORF designation	Gene Product	Gene Category
ECs0002	bifunctional aspartokinase I/homeserine dehydrogenase I	metabolism/housekeeping
ECs0076	isopropylmalate isomerase large subunit	metabolism/housekeeping
ECs0093	cell division protein FtsW	putative function based on homology
ECs0100	UDP-3-O-[3-hydroxymyristoyl] N-acetylglucosamine deacetylase	putative function based on homology
ECs0204	putative transcriptional regulator	putative function based on homology
ECs0406	acetaldehyde dehydrogenase	metabolism/housekeeping
ECs0444	fructokinase	metabolism/housekeeping
ECs0502	putative multidrug transporter membrane\ATP-binding components	putative function based on homology
ECs0523	DNA polymerase III subunits gamma and tau	metabolism/housekeeping
ECs0530	inosine-guanosine kinase	putative function based on homology
ECs0552	putative protease	putative function based on homology
ECs0614	phenylalanine transporter	putative function based on homology
ECs0615	putative transport	putative function based on homology
ECs0647	putative oxidoreductase	putative function based on homology
ECs0745	type II citrate synthase	metabolism/housekeeping
ECs0905	molybdopterin biosynthesis protein MoeA	putative function based on homology
ECs0917	putative dehydrogenase	putative function based on homology
ECs0955	putative dTDP-glucose enzyme	putative function based on homology
ECs0983	putative MFS family transporter protein	putative function based on homology
ECs0998	tetraacyldisaccharide 4'-kinase	metabolism/housekeeping
ECs1044	hypothetical protein	Hypothetical protein
ECs1046	DNA helicase IV	putative function based on homology
ECs1137	cryptic autophosphorylating protein tyrosine kinase Etk	putative function based on homology
ECs1148	hybrid sensory histidine kinase TorS	putative function based on homology
ECs1721	calcium/sodium:proton antiporter	putative function based on homology
ECs1729	nitrate reductase 1 alpha subunit	putative function based on homology
ECs1741	bifunctional acetaldehyde-CoA/alcohol dehydrogenase	putative function based on homology
ECs1833	tryptophan synthase subunit beta	metabolism/housekeeping
ECs1871	peptide transport-like protein	putative function based on homology
ECs1878	putative oxidoreductase	putative function based on homology
ECs2022	glyceraldehyde 3-phosphate dehydrogenase C	putative function based on homology
ECs2045	putative ATP-binding component of a transport system	putative function based on homology
ECs2087	putative ATP-binding component of a transport system	putative function based on homology
ECs2131	glutaminase	metabolism/housekeeping
ECs2320	hypothetical protein	Hypothetical protein
ECs2406	hypothetical protein	Hypothetical protein
ECs2429	6-phosphofructokinase 2	metabolism/housekeeping
ECs2435	part of a kinase	putative function based on homology
ECs2454	bifunctional succinylornithine transaminase/acetylornithine transaminase	putative function based on homology
ECs2488	glyceraldehyde-3-phosphate dehydrogenase	putative function based on homology
ECs2582	biotin sulfoxide reductase 2	putative function based on homology
ECs2583	putative cytochrome C-type protein	putative function based on homology
ECs2589	flagellar biosynthesis protein FlhA	putative function based on homology
ECs3043	DNA-binding transcriptional regulator GalS	putative function based on homology
ECs3059	fructose-specific PTS system IIBC component	putative function based on homology
ECs3076	putative ATP-dependent helicase	putative function based on homology
ECs3076	putative ATP-dependent helicase	putative function based on homology
ECs3207	3-oxoacyl-(acyl carrier protein) synthase I	metabolism/housekeeping
ECs3225	3-ketoacyl-CoA thiolase	metabolism/housekeeping
ECs3228	hypothetical protein	Hypothetical protein
ECs3231	putative prophage Sf6-like integrase	putative function based on homology
ECs3246	multidrug resistance protein Y	putative function based on homology
ECs3300	N-acetylmuramic acid phosphotransfer permease	putative function based on homology
ECs3377	4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase	putative function based on homology
ECs3378	cytoskeletal protein RodZ	putative function based on homology
ECs3464	phospho-2-dehydro-3-deoxyheptonate aldolase	metabolism/housekeeping
ECs3564	D-arabinose 5-phosphate isomerase	putative function based on homology
ECs3639	phosphopyruvate hydratase	metabolism/housekeeping
ECs3676	exonuclease V subunit alpha	putative function based on homology
ECs3746	phenylhydantoinase	metabolism/housekeeping
ECs3751	putative selenate reductase subunit YgfK	putative function based on homology
ECs3773	6-phospho-beta-glucosidase BglA	metabolism/housekeeping
ECs3810	transketolase	putative function based on homology
ECs3833	L-asparaginase II	metabolism/housekeeping
ECs3839	murein transglycosylase C	putative function based on homology
ECs3990	hypothetical protein	Hypothetical protein
ECs3994	putative formate acetyltransferase 3	putative function based on homology

ORF designation	Gene Product	Gene Category
ECs4047	tRNA pseudouridine synthase B	putative function based on homology
ECs4109	malate dehydrogenase	metabolism/housekeeping
ECs4130	sodium/panthothenate symporter	putative function based on homology
ECs4204	putative hydrolase	putative function based on homology
ECs4226	tryptophanyl-tRNA synthetase	metabolism/housekeeping
ECs4259	maltodextrin phosphorylase	putative function based on homology
ECs4265	putative 2-component regulator	putative function based on homology
ECs4359	putative ATP-binding component of a transport system	putative function based on homology
ECs4364	hypothetical protein	Hypothetical protein
ECs4411	endo-1,4-D-glucanase	metabolism/housekeeping
ECs4418	hypothetical protein	Hypothetical protein
ECs4424	peptide ABC transporter	putative function based on homology
ECs4454	periplasmic alpha-amylase precursor	putative function based on homology
ECs4493	putative glycosyl transferase	putative function based on homology
ECs4514	bifunctional phosphopantothenoylcysteine	metabolism/housekeeping
ECs4601	decarboxylase/phosphopantothenate synthase	Hypothetical protein
ECs4689	hypothetical protein	putative function based on homology
ECs4711	potassium transport protein Kup	putative function based on homology
ECs4718	ATP-dependent DNA helicase Rep	putative function based on homology
ECs4841	lipopolysaccharide biosynthesis protein WzzE	putative function based on homology
ECs4861	6-phosphofructokinase	metabolism/housekeeping
ECs4872	DNA-binding transcriptional regulator CytR	putative function based on homology
ECs4920	hypothetical protein	Hypothetical protein
ECs5024	uroporphyrinogen decarboxylase	metabolism/housekeeping
ECs5210	glycerol-3-phosphate acyltransferase	metabolism/housekeeping
ECs5222	UDP-N-acetylglucosamine:L-alanyl-gamma-D-glutamyl-mesodiaminopimelat e ligase	putative function based on homology
	aspartate carbamoyltransferase catalytic subunit	metabolism/housekeeping

ORF designation	Forward Primer	Sequence of Forward Primer (5'-3')	Reverse Primer	Sequence of Reverse Primer (5'-3')	Amplicon size, bp	No. bp analyzed
ECs0002	0002_1558_F	GCATGGCCTAAATCTGGAAA	0002_2214_R	AGCGCAAAACTTTCCCTCA	695	593
ECs0076	0076_42_F	GAAATGGCCGGTCACAGC	0076_682_R	CACCTGACGAAACCACCTT	677	573
ECs0093	0093_293_F	GCGTTTATTCTGGCATCAT	0093_915_R	CACCGACATACCCCAGTTCT	661	560
ECs0100	0100_276_F	GCTTGGGATCGATAACATT	0100_851_R	GTCGTCTGGAAAGGTACAT	614	526
ECs0204	0204_40_F	CTCCGGGAGCTTTTACA	0204_693_R	TCGCGTACAGCTAACCTTG	692	593
ECs0406	0406_61_F	TGGCACCGATCTGATGATTA	0406_676_R	GCCTCAGGCCATTTCATTGAT	654	554
ECs0444	0444_203_F	CTGGCTCAATTTCGCGCTTAC	0444_781_R	AACTCGGAAACCGTTGATA	607	500
ECs0502	0502_115_F	ACTGGTCCCCACAAAGTGG	0502_715_R	TCGAAACGAGCATCAATACG	639	554
ECs0523	0523_843_F	GTATCGAGTGGGAAGCGTTG	0523_1447_R	TCTTTTGTGTCATCACTGG	643	542
ECs0530	0530_238	CGAAGCGTTTATCAGGAAC	0530_899_R	ATGCTGGTTTACGTTTCG	700	607
ECs0552	0552_265_F	GCGCTTCAGCCTGTAAAAAC	0552_885_R	CCCGATTCTCATTTTGTG	659	566
ECs0614	0614_42_F	CGGAAGATACTCGCTCGAAT	0614_621_R	ACAAAATCAGCCCATTCCAG	618	528
ECs0615	0615_140_F	TTCATATTCCAGCCACACCA	0615_769_R	TTTTGATGATCTCGCTGCTG	668	570
ECs0647	0647_31_F	GACTTATCACGGCCACATC	0647_654_R	CAAGCGGTAGGGATGATGA	661	558
ECs0745	0745_309_F	GTGCCCCAGCTCTTCAGAAC	0745_942_R	GACCCGTACATCATGATCC	672	571
ECs0905	0905_574_F	CGCGGATAATCCCTAAGTTG	0905_1187_R	GACACCGCGCTTAATGAGAT	652	545
ECs0917	0917_107_F	CTGCAAGACAAACTCGACCA	0917_763_R	TAGTTGATTCCCCAGGTTGC	695	586
ECs0955	0955_366_F	TGCCACAAAATATTGCCAAA	0955_973_R	TAATCGTTGGTGCAGGTTCA	646	540
ECs0983	0983_281_F	CTTGGCCTGATGATTGGTT	0983_898_R	GCCACCGGATATAGCGTAA	656	571
ECs0998	0998_81_F	GGTTGTATGGCCTGGTGA	0998_673_R	TGTTCAAGCTGAGCAACGTC	631	527
ECs1044	1044_19_F	TTACGCCAACAGGAGTT	1044_594_R	CGATACGCTGATTGGTTGTG	613	523
ECs1046	1046_155_F	ATTCACTGTAAGCGCGGTCT	1046_730_R	CCAAACGCCAGCAATAAAAT	614	516
ECs1137	1137_1518_F	CCGCTTTCTTACTCGTTC	1137_2111_R	ATCGATCTGCTCGTCTGGT	632	540
ECs1148	1148_1491_F	TCCCTGCAATTCTAGCTGTACG	1148_2089_R	TGCAGCAAATGGTGTATGAAT	637	544
ECs1721	1721_338_F	GAAGCCAGTAAAGGCAGACAG	1721_993_R	GGGAAGCAGCCAGACACTAC	694	597
ECs1729	1729_484_F	CGGTGGTTTGTTCGTTCTT	1729_1102_R	TTCCATTCCGGATTGTTTTC	657	569
ECs1741	1741_406_F	TGAATGCGACTGCTGCTGGTC	1741_982_R	ACGAACATCCGGAAACTCAC	615	519
ECs1833	1833_326_F	TTCTTCAATCTGCCGTCTT	1833_898_R	GTCAGGCTTTACTGGCGAAG	611	516
ECs1871	1871_401_F	CTGAAAGCGACCTTACCG	1871_986_R	GGCACTGGACCGTATCAGTT	624	534
ECs1878	1878_393_F	TGTTTGCGATGAATGAT	1878_976_R	ATTTCAKGCCCTTCAACTG	622	517
ECs2022	2022_36_F	TTTCGAGGGTGCATAAG	2022_645_R	CGCGAAGAAGGTGTTGATT	648	552
ECs2045	2045_191_F	GGCGGGGCTATCTATCTT	2045_793_R	AGTTCACCAAGGGGTGTTGAG	641	536
ECs2087	2087_281_F	CACAATCTGCCGAGATA	2087_893_R	TAACGTTACGGGACGTCCA	650	558
ECs2131	2131_76_F	CAAGCGAGTTACCTGCATCA	2131_736_R	CGCAAGAACGTTTTCCATT	699	596
ECs2320	2320_280_F	TCAGAGCGTCATCTCAACG	2320_891_R	CATCAATTGGCCGACTTTT	650	553
ECs2406	2406_587_F	GGCCTCACACCAGAACAGAT	2406_1202_R	CATGATCATTGGCGTACCG	654	564
ECs2429	2429_315_F	AAGCAAGCGGTGAGCAGTAT	2429_882_R	TATCGTCATGGGAGCACAGA	606	503

ORF designation	Forward Primer	Sequence of Forward Primer (5'-3')	Reverse Primer	Sequence of Reverse Primer (5'-3')	Amplicon size, bp	No. bp analyzed
ECs2435	2435_40_F	GAACATCGTGGTGTTCGTTG	2435_666_R	GTTTCATCACCCAGCTTG	665	572
ECs2454	2454_38_F	AACAAAGTGTTCGCAAGCAA	2454_657_R	CTCTACCTGTGCGGTGATTG	658	547
ECs2488	2488_310_F	TGGTCTTCCCTGACTGACG	2488_936_R	AGTAACCGGTTCGTTGTCG	665	561
ECs2582	2582_278_F	GTTCCAGACACGGACCAGAT	2582_887_R	TTTGGAGCGCAATGACTTGAC	648	551
ECs2583	2583_495_F	TCGCCTATGCGAGAACGTTT	2583_1066_R	CGCATTGGGTTATTGTTTTG	611	517
ECs2589	2589_1054_F	CGGCATTATTCTCTGCCATT	2589_1706_R	GTTGGTGGCAATTTCGCTAT	691	589
ECs3043	3043_240_F	TCATCGAACCGATGATTGA	3043_822_R	GGACGTTCTGATGCGTTT	621	530
ECs3059	3059_51_F	GGACGTTTCAAGGACATA	3059_669_R	GCAACTGAAACTGCCCACAGA	657	570
ECs3076	3076a_179_F	GTGCTGGCACACGTTAAAGA	3076a_834_R	GGGTGTCGCCAGTAATCAGT	694	564
ECs3076	3076b_1147_F	CAACGTTCCGGTACAGGTTT	3076b_1722_R	CGACGAAAAGCACCTTCG	612	524
ECs3207	3207_386_F	CATCGCCATCTGCATACAAC	3207_1043_R	CTGGATACCACTGGCCTCAT	696	578
ECs3225	3225_63_F	GCACAGGGGTAACCTAAACC	3225_647_R	GCATGGTCAGACGGAAAAC	623	533
ECs3228	3228_189_F	CCATACGAGGAAACGACGAT	3228_808_R	CGAAATCCTTTGAATGTGG	658	566
ECs3231	3231_570_F	ACCTTGCTGAAGCCATGAAG	3231_1132_R	ACCTTCCCCTCAAGCCAGTC	601	518
ECs3246	3246_302_F	CGACGTACCAACTGATCCTG	3246_923_R	GACTTCACCGGTCAAAATGAA	661	558
ECs3300	3300_365_F	AAATTCCGCCACCATCTTCAC	3300_983_R	GCTGTTGAATCCTTGGCTGT	657	567
ECs3377	3377_470_F	ACCAATGGCGGATTACTG	3377_1098_R	GCATAACCAGGCTCCAATT	667	563
ECs3378	3378_242_F	CCAGCAATCGGCAGTAAGT	3378_836_R	GCCGATCTGCTTCAACATT	633	534
ECs3464	3464_407_F	CGGATTCCCCTGAGTTTGT	3464_1003_R	TAATGACTCCGGCACAACTG	635	546
ECs3564	3564_287_F	CGCGATGTGATGCTGTTTAT	3564_898_R	AGTTTGCCTTTCATCCAC	650	552
ECs3639	3639_69_F	CCCAGAGCTTCTTCGATACG	3639_681_R	AGGCATGAACACTGCTGTTG	651	562
ECs3676	3676_1166_F	ATCGGTCAAGCGTAACGTG	3676_1784_R	GAAGCTGTGGAGCACAAACA	657	574
ECs3746	3746_719_F	CCGCTCTATATCGTCACCT	3746_1302_R	AGGTGCCGTACAGAAAATC	622	530
ECs3751	3751_2250_F	ATGCTGATGGCACCTTAACC	3751_2873_R	CAGGCTGAAAACGGTGATT	662	559
ECs3773	3773_726_F	TGAAAGTCGGCTGTATGCTG	3773_1351_R	ATATCGCCAGTACCGTCTGC	664	519
ECs3810	3810_1293_F	AGGGACGGTTTGTCACTCAC	3810_1932_R	GAGCATGGACGGCAGTACAGA	678	533
ECs3833	3833_299_F	CGCATCTACCAGGGCTTAG	3833_913_R	GTGACTCCGCAACCAAATCT	653	556
ECs3839	3839_474_F	CAAGCAACTTCGCGGATTAT	3839_1047_R	ATTTTGCGCGGTATTCACT	612	319
ECs3990	3990_378_F	CGCGGTAACTGGTTATGGAT	3990_1018_R	GGCTGGAAGTGTGAAAGAC	679	571
ECs3994	3994_524_F	TGGCAGCGCTTTAATTTCT	3994_1136_R	TCGGAAGAATTACCGATTG	651	545
ECs4047	4047_149_F	TGTACGAACCGGTTACCAT	4047_743_R	GCGACGAAGTTTCCAGTA	633	529
ECs4109	4109_39_F	GCCCCAGGGCAGATCTTT	4109_666_R	GGATCGTCCGACCTGTTA	664	572
ECs4130	4130_367_F	CCGCTACCAAGAGTCGCTTC	4130_1002_R	GCAATGGCGCTTAATTGTC	674	584
ECs4204	4204_235_F	ACCTCGTCTGGTGGTTTTC	4204_806_R	AATAATCAGCGTCGGTTTG	610	502
ECs4226	4226_37_F	AACCAATCGCTTCGACACC	4226_671_R	CGCATGACGCAGTTAAAGA	673	574
ECs4259	4259_288_F	TTCAATACTCGCTCCAGCAC	4259_884_R	CAGCAATATCGCAGATCAA	635	533
ECs4265	4265_233_F	GTGGTAGCGTTGAGATTGA	4265_823_R	GCGCTCGTAATAATCCTTC	629	528
ECs4359	4359_780_F	ATCCAGACACCGAGTCCAC	4359_1384_R	TGTTCTGGCAGTTGATGGTC	643	540
ECs4364	4364_20_F	AGCACGACTTCGTCGAA	4364_662_R	AAGATTGCCAACAAATTGG	679	583
ECs4411	4411_428_F	TTTAGCTGCCAGCCTTGT	4411_1047_R	GGCTGCCCTTAGTGTTCAGG	658	541
ECs4418	4418_150_F	CGCTCCTCAATTGGTGT	4418_781_R	TCAATATCGAACATGCGACCA	670	565
ECs4424	4424_906_F	GTATCGATCTGCGTTTGGT	4424_1564_R	GGATGCTGAAGCTTGGCTC	697	599
ECs4454	4454_42_F	TGACACTCCTTCGCTTC	4454_639_R	TTTCCGCCATACCGTCTTA	636	546
ECs4493	4493_361_F	GCAACCGACTCACTGACGTA	4493_956_R	TATAATCGGGCGATGATT	634	544
ECs4514	4514_239_F	GGCCATATTGAGCTGGTAA	4514_864_R	TGGCCTGCTTTTGATTT	664	547
ECs4601	4601_38_F	CGACTGCCACACCAAGAGTA	4601_602_R	TTTTGCTCACCGCTATTTCG	603	518
ECs4689	4689_322_F	GCTGGTGATTATGGGCTAA	4689_979_R	TCCATTTCGGAGGTGTAAT	696	547
ECs4711	4711_22_F	GTCTAAACCCGGCCAAAC	4711_654_R	CATACTGGCTGGTGTGGTG	669	572
ECs4718	4718_366_F	GAGAGTTCTGGCTGCAAAACC	4718_965_R	AATCATCAGGAAGGCACGAC	638	518
ECs4841	4841_181_F	CCGTTACAGCGTTCTGACA	4841_786_R	CCATACGGGAAGCCAGAATA	644	555
ECs4861	4861_177_F	TCACAAAATTGCGTCAGGTC	4861_829_R	GCAACGTCAAGCGTAATGAA	691	595
ECs4872	4872_234_F	CCTTAGCCATTGCCCTGTTA	4872_828_R	GCACAATCAGCACAAATACCG	633	543
ECs4920	4920_376_F	TGAACTGGGTACGTGATGA	4920_965_R	GCCGTGACCAAGGTTAAAGA	628	541
ECs5024	5024_1136_F	GGCGTAAGTACCCACTTCA	5024_1730_R	ATTGCCAAAGCGGTAGAAGA	633	515
ECs5210	5210_149_F	AAGCAAGGCATTGAGCTGAT	5210_772_R	TCCAGCAAAACTTCCCATTC	662	566
ECs5222	5222_284_F	GTCAACTTCCGCCATCACTT	5222_913_R	TGGCTTAATCCGCTATATCAGAAA	671	575

Technical Appendix Table 2. SNP list of all 50 strains investigated

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs0076.SNP-1212	ECs0076.SNP-1164	ECs0093.SNP-687	ECs0100.SNP-406	ECs0204.SNP-573	ECs0406.SNP-494
493/89	C	C	C	G	C	C
86-24	C	T	C	G	C	C
87-14	C	T	C	G	C	C
CB9615	T	C	C	A	C	T
EC4115	C	T	C	G	C	C
EDL933	C	T	C	G	C	C
LSU-61	C	C	C	G	C	C
Sakai	C	T	C	G	C	C
TB182A	T	C	C	A	C	C
TW14359	C	T	C	G	C	C
TW14588	C	T	C	G	C	C
SNPO157_01	C	T	C	G	C	C
SNPO157_02	C	T	T	G	T	C
SNPO157_03	C	C	C	G	C	C
SNPO157_04	C	T	C	G	C	C
SNPO157_05	C	T	C	G	C	C
SNPO157_06	C	T	C	G	C	C
SNPO157_07	C	T	T	G	C	C
SNPO157_08	C	T	C	G	C	C
SNPO157_09	C	C	C	G	C	C
SNPO157_10	C	T	C	G	C	C
SNPO157_11	C	C	C	G	C	C
SNPO157_12	C	C	C	G	C	C
SNPO157_13	C	T	T	G	C	C
SNPO157_14	C	T	C	G	C	C
SNPO157_15	C	T	C	G	C	C
SNPO157_16	C	T	C	G	C	C
SNPO157_17	C	T	C	G	C	C
SNPO157_18	C	T	C	G	C	C
SNPO157_19	C	T	C	G	C	C
SNPO157_20	C	T	C	G	C	C
SNPO157_21	C	C	C	G	C	C
SNPO157_22	C	T	C	G	C	C
SNPO157_23	C	T	C	G	C	C
SNPO157_24	C	T	C	G	C	C
SNPO157_25	C	T	C	G	C	C
SNPO157_26	C	T	C	G	C	C
SNPO157_27	C	T	C	G	C	C
SNPO157_28	C	T	C	G	C	C
SNPO157_29	C	T	C	G	C	C
SNPO157_30	C	T	C	G	C	C
SNPO157_31	C	T	C	G	C	C
SNPO157_32	C	T	C	G	C	C
SNPO157_33	C	T	C	G	C	C
SNPO157_34	C	C	C	G	C	C
SNPO157_35	C	C	C	G	C	C
SNPO157_36	C	C	C	G	C	C
SNPO157_37	C	C	C	G	C	C
SNPO157_38	C	T	C	G	C	C
SNPO157_39	C	C	C	G	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs0444.SNP-260	ECs0502.SNP-342	ECs0523.1134	ECs0552.SNP-186	ECs0614.SNP-138	ECs0614.SNP-283	ECs0647.SNP-216
493/89	C	T	C	C	C	A	C
86-24	C	T	C	C	C	T	T
87-14	C	T	C	C	T	T	T
CB9615	C	G	C	C	C	T	C
EC4115	C	T	C	C	C	T	T
EDL933	C	T	C	C	C	T	T
LSU-61	C	T	T	C	C	T	C
Sakai	C	T	C	C	C	T	T
TB182A	C	G	C	C	C	T	C
TW14359	C	T	C	C	C	T	T
TW14588	C	T	C	C	C	T	T

Strain ID	Locus tag (ORF designation) SNP position within ORF							
	ECs0444.SNP-		ECs0502.SNP-		ECs0552.SNP-		ECs0614.SNP-	
	260	342	ECs0523.1134	186	138	283	216	
SNPO157_01	C	T	C	C	T	T	T	T
SNPO157_02	C	T	C	C	C	T	T	T
SNPO157_03	C	T	C	C	C	A	C	
SNPO157_04	C	T	C	C	T	T	T	T
SNPO157_05	C	T	C	T	C	T	T	T
SNPO157_06	C	T	C	C	C	T	T	T
SNPO157_07	C	T	C	C	C	T	T	T
SNPO157_08	C	T	C	C	T	T	T	T
SNPO157_09	C	T	C	C	C	A	C	
SNPO157_10	C	T	C	C	T	T	T	T
SNPO157_11	C	T	C	C	C	A	C	
SNPO157_12	C	T	C	C	C	T	C	
SNPO157_13	C	T	C	C	T	T	T	
SNPO157_14	C	T	C	C	T	T	T	
SNPO157_15	C	T	C	C	T	T	T	
SNPO157_16	C	T	C	C	T	T	T	
SNPO157_17	C	T	C	C	T	T	T	
SNPO157_18	C	T	C	C	T	T	T	
SNPO157_19	C	T	C	C	T	T	T	
SNPO157_20	C	T	C	C	C	T	T	
SNPO157_21	C	T	C	C	C	A	C	
SNPO157_22	C	T	C	C	C	T	T	
SNPO157_23	C	T	C	C	C	T	T	
SNPO157_24	T	T	C	C	T	T	T	
SNPO157_25	C	T	C	C	C	T	T	
SNPO157_26	C	T	C	C	T	T	T	
SNPO157_27	C	T	C	C	T	T	T	
SNPO157_28	C	T	C	C	C	T	T	
SNPO157_29	C	T	C	C	T	T	T	
SNPO157_30	C	T	C	C	C	T	T	
SNPO157_31	C	T	C	C	C	T	T	
SNPO157_32	C	T	C	C	C	T	T	
SNPO157_33	C	T	C	C	C	T	T	
SNPO157_34	C	T	C	C	C	A	C	
SNPO157_35	C	T	C	C	C	A	C	
SNPO157_36	C	T	C	C	C	T	C	
SNPO157_37	C	T	C	C	C	A	C	
SNPO157_38	C	T	C	C	C	T	T	
SNPO157_39	C	T	C	C	C	A	C	

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs0647.SNP-		ECs0647.SNP-		ECs0905.SNP-	
	306	556	453	217	578	745
493/89	C	C	C	C	C	C
86-24	C	C	C	G	C	C
87-14	C	C	C	G	C	C
CB9615	C	C	T	G	C	C
EC4115	T	C	C	GG	C	C
EDL933	C	C	C	GG	C	C
LSU-61	C	C	C	GG	C	A
Sakai	C	C	C	GG	C	C
TB182A	C	C	T	GG	C	C
TW14359	T	C	C	GG	C	C
TW14588	C	C	C	GG	C	C
SNPO157_01	C	C	C	GG	C	C
SNPO157_02	T	C	C	GG	C	C
SNPO157_03	C	C	C	GG	C	C
SNPO157_04	C	C	C	GG	C	C
SNPO157_05	C	C	C	GG	C	C
SNPO157_06	C	C	C	GG	C	C
SNPO157_07	T	C	C	GG	C	C
SNPO157_08	C	C	C	GG	C	C
SNPO157_09	C	C	C	GG	C	C
SNPO157_10	C	C	C	GG	C	C
SNPO157_11	C	C	C	GG	C	C
SNPO157_12	C	C	C	GG	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs0647.SNP-306	ECs0647.SNP-556	ECs0905.SNP-453	ECs0905.SNP-217	ECs0917.SNP-578	ECs0955.SNP-745
SNPO157_13	T	C	C	G	C	C
SNPO157_14	C	C	C	G	C	C
SNPO157_15	C	C	C	G	C	C
SNPO157_16	C	C	C	GG	C	C
SNPO157_17	C	C	C	GG	C	C
SNPO157_18	C	C	C	GG	C	C
SNPO157_19	C	C	C	GG	C	C
SNPO157_20	C	C	C	GC	C	C
SNPO157_21	C	C	C	GG	C	C
SNPO157_22	C	C	C	GG	C	C
SNPO157_23	C	C	C	GG	C	C
SNPO157_24	C	C	C	GG	C	C
SNPO157_25	C	C	C	GG	C	C
SNPO157_26	C	C	C	G	C	C
SNPO157_27	C	C	C	GG	C	C
SNPO157_28	C	C	C	GG	T	C
SNPO157_29	C	C	C	GG	C	C
SNPO157_30	C	C	C	GG	C	C
SNPO157_31	C	C	C	GG	T	C
SNPO157_32	C	C	C	GG	C	C
SNPO157_33	C	C	C	G	T	C
SNPO157_34	C	C	C	C	C	C
SNPO157_35	C	C	C	C	C	C
SNPO157_36	C	G	C	G	C	C
SNPO157_37	C	C	C	C	C	C
SNPO157_38	C	C	C	G	C	C
SNPO157_39	C	C	C	C	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs0955.SNP-719	ECs0998.SNP-375	ECs1044.SNP-2080	ECs1044.SNP-1773	ECs1046.SNP-319	ECs1046.SNP-358
493/89	C	C	A	C	C	C
86-24	C	T	A	C	C	C
87-14	C	C	A	C	C	C
CB9615	C	C	A	C	C	A
EC4115	C	C	G	C	C	C
EDL933	C	C	A	T	C	C
LSU-61	C	C	A	C	C	C
Sakai	C	C	A	C	C	C
TB182A	T	C	A	C	C	A
TW14359	C	C	G	C	C	C
TW14588	C	C	A	C	C	C
SNPO157_01	C	C	A	C	C	C
SNPO157_02	C	C	A	C	C	C
SNPO157_03	C	C	A	C	C	C
SNPO157_04	C	C	A	C	C	C
SNPO157_05	C	C	A	C	C	C
SNPO157_06	C	C	A	C	C	C
SNPO157_07	C	C	A	C	C	C
SNPO157_08	C	C	A	C	C	C
SNPO157_09	C	C	A	C	C	C
SNPO157_10	C	C	A	C	C	C
SNPO157_11	C	C	A	C	C	C
SNPO157_12	C	C	A	C	C	C
SNPO157_13	C	C	A	C	C	C
SNPO157_14	C	C	A	C	C	C
SNPO157_15	C	C	A	C	C	C
SNPO157_16	C	C	A	C	C	C
SNPO157_17	C	C	A	C	C	C
SNPO157_18	C	C	A	C	C	C
SNPO157_19	C	C	A	C	C	C
SNPO157_20	C	C	A	C	C	C
SNPO157_21	C	C	A	C	C	C
SNPO157_22	C	C	A	C	C	C
SNPO157_23	C	C	A	C	C	C
SNPO157_24	C	C	A	C	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs0955.SNP-719	ECs0998.SNP-375	ECs1044.SNP-2080	ECs1044.SNP-1773	ECs1046.SNP-319	ECs1046.SNP-358
SNPO157_25	C	C	A	C	C	C
SNPO157_26	C	C	A	C	C	C
SNPO157_27	C	C	A	C	C	C
SNPO157_28	C	C	A	C	T	C
SNPO157_29	C	C	A	C	C	C
SNPO157_30	C	C	A	C	C	C
SNPO157_31	C	C	A	C	T	C
SNPO157_32	C	C	A	C	C	C
SNPO157_33	C	C	A	C	T	C
SNPO157_34	C	C	A	C	C	C
SNPO157_35	C	C	A	C	C	C
SNPO157_36	C	C	A	C	C	C
SNPO157_37	C	C	A	C	C	C
SNPO157_38	C	C	A	C	C	C
SNPO157_39	C	C	A	C	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs1137.SNP-524	ECs1137.SNP-230	ECs1148.SNP-1014	ECs1148.SNP-940	ECs1729.SNP-786	ECs1741.SNP-2155
493/89	A	C	C	A	C	A
86-24	A	C	C	G	C	G
87-14	A	C	C	G	C	G
CB9615	G	C	C	G	C	G
EC4115	A	C	C	G	T	G
EDL933	A	C	C	GG	CC	G
LSU-61	A	T	C	GG	CC	G
Sakai	A	C	C	G	C	G
TB182A	G	C	A	G	C	G
TW14359	A	C	C	GG	TC	G
TW14588	A	C	C	G	C	G
SNPO157_01	A	C	C	G	C	G
SNPO157_02	A	C	C	G	T	G
SNPO157_03	A	C	C	A	C	A
SNPO157_04	A	C	C	GG	CC	G
SNPO157_05	A	C	C	G	CC	G
SNPO157_06	A	C	C	GG	CC	G
SNPO157_07	A	C	C	GG	TC	G
SNPO157_08	A	C	C	G	C	G
SNPO157_09	A	C	C	A	CC	G
SNPO157_10	A	C	C	G	CC	G
SNPO157_11	A	C	C	A	CC	A
SNPO157_12	A	C	C	A	CT	A
SNPO157_13	A	C	C	G	TC	G
SNPO157_14	A	C	C	GG	CC	G
SNPO157_15	A	C	C	GG	CC	G
SNPO157_16	A	C	C	G	CC	G
SNPO157_17	A	C	C	GG	CC	G
SNPO157_18	A	C	C	GG	CC	G
SNPO157_19	A	C	C	GG	CC	G
SNPO157_20	A	C	C	GG	CC	G
SNPO157_21	A	C	C	A	CC	A
SNPO157_22	A	C	C	G	CC	G
SNPO157_23	A	C	C	G	CC	G
SNPO157_24	A	C	C	G	CC	G
SNPO157_25	A	C	C	G	CC	G
SNPO157_26	A	C	C	GG	CC	G
SNPO157_27	A	C	C	GG	CC	G
SNPO157_28	A	C	C	GG	CC	G
SNPO157_29	A	C	C	GG	CC	G
SNPO157_30	A	C	C	GGG	CC	G
SNPO157_31	A	C	C	GG	CC	G
SNPO157_32	A	C	C	G	CC	G
SNPO157_33	A	C	C	G	CC	G
SNPO157_34	A	C	C	A	CC	A
SNPO157_35	A	C	C	A	CC	A

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs1137.SNP-524	ECs1137.SNP-230	ECs1148.SNP-1014	ECs1148.SNP-940	ECs1729.SNP-786	ECs1741.SNP-2155
SNPO157_36	A	C	C	G	C	G
SNPO157_37	A	C	C	A	C	A
SNPO157_38	A	C	C	G	C	G
SNPO157_39	A	C	C	A	C	A

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs1833.SNP-474	ECs1871.SNP-1109	ECs1871.SNP-978	ECs1878.SNP-829	ECs2022.SNP-793	ECs2045.SNP-677
493/89	C	G	A	T	T	C
86-24	C	A	G	G	T	C
87-14	C	A	G	T	T	C
CB9615	C	G	G	T	T	T
EC4115	C	A	G	T	T	C
EDL933	C	A	G	G	T	C
LSU-61	C	G	G	T	T	C
Sakai	C	A	G	G	T	C
TB182A	C	G	G	T	T	T
TW14359	C	A	G	T	T	C
TW14588	C	A	G	G	T	C
SNPO157_01	C	A	G	T	T	C
SNPO157_02	C	A	G	T	T	C
SNPO157_03	C	G	A	T	T	C
SNPO157_04	C	A	G	T	T	C
SNPO157_05	C	A	G	G	T	C
SNPO157_06	C	A	G	G	T	C
SNPO157_07	C	A	G	T	T	C
SNPO157_08	C	A	G	T	T	C
SNPO157_09	C	G	A	T	T	C
SNPO157_10	C	A	G	T	T	C
SNPO157_11	C	G	A	T	T	C
SNPO157_12	C	G	A	T	T	C
SNPO157_13	C	A	G	T	T	C
SNPO157_14	C	A	G	T	T	C
SNPO157_15	C	A	G	T	T	C
SNPO157_16	C	A	G	G	T	C
SNPO157_17	C	A	G	T	T	C
SNPO157_18	C	A	G	G	T	C
SNPO157_19	C	A	G	T	T	C
SNPO157_20	C	A	G	G	T	C
SNPO157_21	T	G	A	T	T	C
SNPO157_22	C	A	G	G	T	C
SNPO157_23	C	A	G	G	T	C
SNPO157_24	C	A	G	T	T	C
SNPO157_25	C	A	G	G	T	C
SNPO157_26	C	A	G	T	T	C
SNPO157_27	C	A	G	G	T	C
SNPO157_28	C	A	G	G	T	C
SNPO157_29	C	A	G	T	T	C
SNPO157_30	C	A	G	G	T	C
SNPO157_31	C	A	G	G	T	C
SNPO157_32	C	A	G	G	T	C
SNPO157_33	C	A	G	G	T	C
SNPO157_34	C	G	A	T	T	C
SNPO157_35	C	G	A	T	T	C
SNPO157_36	C	A	G	T	A	C
SNPO157_37	C	G	A	T	T	C
SNPO157_38	C	A	G	G	T	C
SNPO157_39	C	G	A	T	T	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs2087.SNP-253	ECs2320.SNP-486	ECs2320.SNP-719	ECs2435.SNP-123	ECs2435.SNP-458	ECs2454.SNP-1064
493/89	C	G	A	G	A	G
86-24	T	A	A	G	T	G

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs2087.SNP-253	ECs2320.SNP-486	ECs2320.SNP-719	ECs2435.SNP-123	ECs2435.SNP-458	ECs2454.SNP-1064
87-14	T	A	T	G	A	A
CB9615	C	G	A	G	A	G
EC4115	T	A	T	A	A	G
EDL933	T	A	A	G	T	G
LSU-61	C	G	A	G	A	G
Sakai	T	A	A	G	T	G
TB182A	C	G	A	G	A	G
TW14359	T	A	T	A	A	G
TW14588	T	A	A	G	T	G
SNPO157_01	T	A	T	G	A	A
SNPO157_02	T	A	T	G	A	G
SNPO157_03	C	G	A	G	A	G
SNPO157_04	T	A	T	G	A	A
SNPO157_05	T	A	A	G	A	G
SNPO157_06	T	A	A	G	A	G
SNPO157_07	T	A	T	G	A	G
SNPO157_08	T	A	T	G	A	A
SNPO157_09	C	G	A	G	A	G
SNPO157_10	T	A	T	G	A	A
SNPO157_11	C	G	A	G	A	G
SNPO157_12	C	G	A	G	A	G
SNPO157_13	T	A	T	G	A	G
SNPO157_14	T	A	T	G	A	A
SNPO157_15	T	A	T	G	A	A
SNPO157_16	T	A	T	G	A	A
SNPO157_17	T	A	T	G	A	A
SNPO157_18	T	A	A	G	A	G
SNPO157_19	T	A	T	G	A	A
SNPO157_20	T	A	A	G	T	G
SNPO157_21	C	G	A	G	A	G
SNPO157_22	T	A	A	G	A	G
SNPO157_23	T	A	A	G	T	G
SNPO157_24	T	A	T	G	A	A
SNPO157_25	T	A	A	G	A	G
SNPO157_26	T	A	T	G	A	A
SNPO157_27	T	A	A	G	A	G
SNPO157_28	T	A	A	G	A	G
SNPO157_29	T	A	T	G	A	A
SNPO157_30	T	A	A	G	A	G
SNPO157_31	T	A	A	G	A	G
SNPO157_32	T	A	A	G	A	G
SNPO157_33	T	A	A	G	A	G
SNPO157_34	C	G	A	G	A	G
SNPO157_35	C	G	A	G	A	G
SNPO157_36	T	G	A	G	A	G
SNPO157_37	C	G	A	G	A	G
SNPO157_38	T	A	A	G	A	G
SNPO157_39	C	G	A	G	A	G

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs2454.SNP-693	ECs2582.SNP-2038	ECs2583.SNP-73	ECs2589.SNP-804	ECs2589.SNP-771	ECs3043.ESNP-721
493/89	A	G	G	A	A	C
86-24	A	G	T	A	A	C
87-14	A	A	T	A	A	C
CB9615	G	G	G	A	C	C
EC4115	A	G	T	A	A	C
EDL933	A	G	T	A	A	C
LSU-61	A	G	G	C	A	C
Sakai	A	G	T	A	A	C
TB182A	G	G	G	A	C	C
TW14359	A	G	T	A	A	C
TW14588	A	G	T	A	A	C
SNPO157_01	A	G	T	A	A	C
SNPO157_02	A	G	T	A	A	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs2454.SNP-693	ECs2582.SNP-2038	ECs2583.SNP-73	ECs2589.SNP-804	ECs2589.SNP-771	ECs3043.ESNP-721
SNPO157_03	A	G	G	A	A	C
SNPO157_04	A	G	T	A	A	C
SNPO157_05	A	G	T	A	A	C
SNPO157_06	A	G	T	A	A	C
SNPO157_07	A	G	T	A	A	C
SNPO157_08	A	G	T	A	A	C
SNPO157_09	A	G	G	A	A	C
SNPO157_10	A	G	T	A	A	C
SNPO157_11	A	G	G	A	A	C
SNPO157_12	A	G	G	A	A	C
SNPO157_13	A	G	T	A	A	C
SNPO157_14	A	G	T	A	A	C
SNPO157_15	A	G	T	A	A	C
SNPO157_16	A	G	T	A	A	C
SNPO157_17	A	G	T	A	A	C
SNPO157_18	A	G	T	A	A	C
SNPO157_19	A	G	T	A	A	C
SNPO157_20	A	G	T	A	A	C
SNPO157_21	A	G	G	A	A	C
SNPO157_22	A	G	T	A	A	C
SNPO157_23	A	G	T	A	A	C
SNPO157_24	A	G	T	A	A	T
SNPO157_25	A	G	T	A	A	C
SNPO157_26	A	G	T	A	A	C
SNPO157_27	A	G	T	A	A	C
SNPO157_28	A	G	T	A	A	C
SNPO157_29	A	G	T	A	A	C
SNPO157_30	A	G	T	A	A	C
SNPO157_31	A	G	T	A	A	C
SNPO157_32	A	G	T	A	A	C
SNPO157_33	A	G	T	A	A	C
SNPO157_34	A	G	G	A	A	C
SNPO157_35	A	G	G	A	A	C
SNPO157_36	A	G	T	A	A	C
SNPO157_37	A	G	G	A	A	C
SNPO157_38	A	G	T	A	A	C
SNPO157_39	A	G	G	A	A	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3059.SNP-1440	ECs3076.SNP-311	ECs3076.SNP-573	ECs3076.SNP-1413	ECs3076.SNP-1689	ECs3207.SNP-291
493/89	C	T	T	G	G	C
86-24	A	T	T	G	G	C
87-14	A	C	T	G	A	C
CB9615	C	T	T	G	G	A
EC4115	A	T	T	G	G	C
EDL933	A	T	T	G	G	C
LSU-61	C	T	T	G	G	C
Sakai	A	T	T	G	G	C
TB182A	C	T	G	A	G	A
TW14359	A	T	T	G	G	C
TW14588	A	T	T	G	G	C
SNPO157_01	A	C	T	G	A	C
SNPO157_02	A	T	T	G	G	C
SNPO157_03	C	T	T	G	G	C
SNPO157_04	A	C	T	G	A	C
SNPO157_05	A	T	T	G	G	C
SNPO157_06	A	T	T	G	G	C
SNPO157_07	A	T	T	G	G	C
SNPO157_08	A	T	T	G	G	C
SNPO157_09	C	T	T	G	G	C
SNPO157_10	A	C	T	G	A	C
SNPO157_11	C	T	T	G	G	C
SNPO157_12	C	T	T	G	G	C
SNPO157_13	A	T	T	G	G	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3059.SNP-1440	ECs3076.SNP-311	ECs3076.SNP-573	ECs3076.SNP-1413	ECs3076.SNP-1689	ECs3207.SNP-291
SNPO157_14	A	C	T	G	A	C
SNPO157_15	A	C	T	G	A	C
SNPO157_16	A	C	T	G	A	C
SNPO157_17	A	C	T	G	A	C
SNPO157_18	A	T	T	G	G	C
SNPO157_19	A	C	T	G	A	C
SNPO157_20	A	T	T	G	G	C
SNPO157_21	C	T	T	G	G	C
SNPO157_22	A	T	T	G	G	C
SNPO157_23	A	T	T	G	G	C
SNPO157_24	A	C	T	G	A	C
SNPO157_25	A	T	T	G	G	C
SNPO157_26	A	C	T	G	A	C
SNPO157_27	A	T	T	G	G	C
SNPO157_28	A	T	T	G	G	C
SNPO157_29	A	C	T	G	A	C
SNPO157_30	A	T	T	G	G	C
SNPO157_31	A	T	T	G	G	C
SNPO157_32	A	T	T	G	G	C
SNPO157_33	A	T	T	G	G	C
SNPO157_34	C	T	T	G	G	C
SNPO157_35	C	T	T	G	G	C
SNPO157_36	A	T	T	G	G	C
SNPO157_37	C	T	T	G	G	C
SNPO157_38	A	T	T	G	G	C
SNPO157_39	C	T	T	G	G	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3225.SNP-1098	ECs3225.SNP-904	ECs3231.SNP-1105	ECs3246.SNP-1113	ECs3246.SNP-743	ECs3300.SNP-458
493/89	T	G	C	C	G	G
86-24	C	G	C	C	A	G
87-14	C	G	C	C	A	G
CB9615	T	G	T	T	G	G
EC4115	C	G	C	C	A	A
EDL933	C	G	C	C	A	G
LSU-61	T	G	C	C	G	G
Sakai	C	G	C	C	A	G
TB182A	T	G	T	T	G	G
TW14359	C	G	C	C	A	G
TW14588	C	G	C	C	A	G
SNPO157_01	C	G	C	C	A	G
SNPO157_02	C	G	C	C	A	G
SNPO157_03	T	G	C	C	G	G
SNPO157_04	C	G	C	C	A	G
SNPO157_05	C	G	C	C	A	G
SNPO157_06	C	G	C	C	A	G
SNPO157_07	C	G	C	C	A	G
SNPO157_08	C	G	C	C	A	G
SNPO157_09	T	G	C	C	G	G
SNPO157_10	C	G	C	C	A	G
SNPO157_11	T	G	C	C	G	G
SNPO157_12	T	G	C	C	G	G
SNPO157_13	C	G	C	C	A	G
SNPO157_14	C	G	C	C	A	G
SNPO157_15	C	G	C	C	A	G
SNPO157_16	C	G	C	C	A	G
SNPO157_17	C	G	C	C	A	G
SNPO157_18	C	G	C	C	A	G
SNPO157_19	C	G	C	C	A	G
SNPO157_20	C	G	C	C	A	G
SNPO157_21	T	G	C	C	G	G
SNPO157_22	C	G	C	C	A	G
SNPO157_23	C	G	C	C	A	G
SNPO157_24	C	G	C	C	A	G

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3225.SNP-1098	ECs3225.SNP-904	ECs3231.SNP-1105	ECs3246.SNP-1113	ECs3246.SNP-743	ECs3300.SNP-458
SNPO157_25	C	G	C	C	A	G
SNPO157_26	C	G	C	C	A	G
SNPO157_27	C	G	C	C	A	G
SNPO157_28	C	G	C	C	A	G
SNPO157_29	C	G	C	C	A	G
SNPO157_30	C	G	C	C	A	G
SNPO157_31	C	G	C	C	A	G
SNPO157_32	C	G	C	C	A	G
SNPO157_33	C	G	C	C	A	G
SNPO157_34	T	G	C	C	G	G
SNPO157_35	T	G	C	C	G	G
SNPO157_36	C	A	C	C	A	G
SNPO157_37	T	G	C	C	G	G
SNPO157_38	C	G	C	C	A	G
SNPO157_39	T	G	C	C	G	G

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3300.SNP-855	ECs3377.SNP-522	ECs3377.SNP-226	ECs3377.SNP-127	ECs3464.SNP-297	ECs3464.SNP-227
493/89	T	A	T	C	T	C
86-24	T	A	T	C	G	C
87-14	T	A	T	C	G	C
CB9615	T	A	G	C	G	C
EC4115	T	A	T	C	G	C
EDL933	T	A	T	C	G	C
LSU-61	T	A	T	T	G	C
Sakai	T	A	T	C	G	C
TB182A	A	A	G	C	G	C
TW14359	T	A	T	C	G	C
TW14588	T	A	T	C	G	C
SNPO157_01	T	A	T	C	G	C
SNPO157_02	T	A	T	C	G	C
SNPO157_03	T	A	T	C	T	C
SNPO157_04	T	A	T	C	G	C
SNPO157_05	T	A	T	C	G	C
SNPO157_06	T	A	T	C	G	C
SNPO157_07	T	A	T	C	G	C
SNPO157_08	T	C	T	C	G	C
SNPO157_09	T	A	T	C	T	C
SNPO157_10	T	A	T	C	G	C
SNPO157_11	T	A	T	C	T	C
SNPO157_12	T	A	T	C	T	C
SNPO157_13	T	A	T	C	G	C
SNPO157_14	T	A	T	C	G	C
SNPO157_15	T	A	T	C	G	C
SNPO157_16	T	A	T	C	G	C
SNPO157_17	T	A	T	C	G	C
SNPO157_18	T	A	T	C	G	C
SNPO157_19	T	A	T	C	G	C
SNPO157_20	T	A	T	C	G	C
SNPO157_21	T	A	T	C	T	C
SNPO157_22	T	A	T	C	G	C
SNPO157_23	T	A	T	C	G	C
SNPO157_24	T	A	T	C	G	C
SNPO157_25	T	A	T	C	G	C
SNPO157_26	T	A	T	C	G	C
SNPO157_27	T	A	T	C	G	C
SNPO157_28	T	A	T	C	G	C
SNPO157_29	T	A	T	C	G	C
SNPO157_30	T	A	T	C	G	C
SNPO157_31	T	A	T	C	G	C
SNPO157_32	T	A	T	C	G	A
SNPO157_33	T	A	T	C	G	C
SNPO157_34	T	A	T	C	G	C
SNPO157_35	T	A	T	C	T	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3300.SNP-855	ECs3377.SNP-522	ECs3377.SNP-226	ECs3377.SNP-127	ECs3464.SNP-297	ECs3464.SNP-227
SNPO157_36	T	A	T	C	G	C
SNPO157_37	T	A	T	C	T	C
SNPO157_38	T	A	T	C	G	C
SNPO157_39	T	A	T	C	T	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3564.SNP-786	ECs3676.SNP-147	ECs3746.SNP-984	ECs3746.SNP-1136	ECs3773.SNP-1100	ECs3810.SNP-429
493/89	C	C	T	A	A	A
86-24	C	C	C	A	A	G
87-14	A	C	C	A	A	G
CB9615	C	T	C	A	C	G
EC4115	A	C	C	A	A	G
EDL933	C	C	C	A	A	G
LSU-61	C	C	C	A	A	G
Sakai	C	C	C	A	A	G
TB182A	C	T	C	A	C	G
TW14359	A	C	C	A	A	G
TW14588	C	C	C	A	A	G
SNPO157_01	A	C	C	A	A	G
SNPO157_02	A	C	C	A	A	G
SNPO157_03	C	C	T	A	A	A
SNPO157_04	A	C	C	A	A	G
SNPO157_05	C	C	C	A	A	G
SNPO157_06	C	C	C	A	A	G
SNPO157_07	A	C	C	A	A	G
SNPO157_08	A	C	C	A	A	G
SNPO157_09	C	C	T	A	A	A
SNPO157_10	A	C	C	A	A	G
SNPO157_11	C	C	T	A	A	A
SNPO157_12	C	C	T	A	A	A
SNPO157_13	A	C	C	A	A	G
SNPO157_14	A	C	C	A	A	G
SNPO157_15	A	C	C	A	A	G
SNPO157_16	A	C	C	A	A	G
SNPO157_17	A	C	C	A	A	G
SNPO157_18	C	C	C	A	A	G
SNPO157_19	A	C	C	A	A	G
SNPO157_20	C	C	C	A	A	G
SNPO157_21	C	C	T	A	A	A
SNPO157_22	C	C	C	A	A	G
SNPO157_23	C	C	C	A	A	G
SNPO157_24	A	C	C	A	A	G
SNPO157_25	C	C	C	C	A	G
SNPO157_26	A	C	C	A	A	G
SNPO157_27	C	C	C	A	A	G
SNPO157_28	C	C	C	A	A	G
SNPO157_29	A	C	C	A	A	G
SNPO157_30	C	C	C	A	A	G
SNPO157_31	C	C	C	A	A	G
SNPO157_32	C	C	C	C	A	G
SNPO157_33	C	C	C	A	A	G
SNPO157_34	C	C	T	A	A	A
SNPO157_35	C	C	T	A	A	A
SNPO157_36	C	C	C	A	A	G
SNPO157_37	C	C	T	A	A	A
SNPO157_38	C	C	C	A	A	G
SNPO157_39	C	C	T	A	A	A

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3833.SNP-365	ECs3839.SNP-793	ECs3994.SNP-1663	ECs4047.SNP-272	ECs4109.SNP-873	ECs4109.SNP-812
493/89	T	G	C	G	C	T
86-24	T	G	T	G	C	C

Strain ID	Locus tag (ORF designation) SNP position within ORF					
	ECs3833.SNP-365	ECs3839.SNP-793	ECs3994.SNP-1663	ECs4047.SNP-272	ECs4109.SNP-873	ECs4109.SNP-812
87-14	T	G	T	G	C	C
CB9615	T	A	C	G	A	C
EC4115	T	G	T	A	C	C
EDL933	T	G	T	G	C	C
LSU-61	T	G	C	G	C	C
Sakai	T	G	T	G	C	C
TB182A	C	A	C	G	A	C
TW14359	T	G	T	A	C	C
TW14588	T	G	T	G	C	C
SNPO157_01	T	G	T	G	C	C
SNPO157_02	T	G	T	G	C	C
SNPO157_03	T	G	C	G	C	T
SNPO157_04	T	G	T	G	C	C
SNPO157_05	T	G	T	G	C	C
SNPO157_06	T	G	T	G	C	C
SNPO157_07	T	G	T	G	C	C
SNPO157_08	T	G	T	G	C	C
SNPO157_09	T	G	C	G	C	T
SNPO157_10	T	G	T	G	C	C
SNPO157_11	T	G	C	G	C	T
SNPO157_12	T	G	C	G	C	T
SNPO157_13	T	G	T	G	C	C
SNPO157_14	T	G	T	G	C	C
SNPO157_15	T	G	T	G	C	C
SNPO157_16	T	G	T	G	C	C
SNPO157_17	T	G	T	G	C	C
SNPO157_18	T	G	T	G	C	C
SNPO157_19	T	G	T	G	C	C
SNPO157_20	T	G	T	G	C	C
SNPO157_21	T	G	C	G	C	T
SNPO157_22	T	G	T	G	C	C
SNPO157_23	T	G	T	G	C	C
SNPO157_24	T	G	T	G	C	C
SNPO157_25	T	G	T	G	C	C
SNPO157_26	T	G	T	G	C	C
SNPO157_27	T	G	T	G	C	C
SNPO157_28	T	G	T	G	C	C
SNPO157_29	T	G	T	G	C	C
SNPO157_30	T	G	T	G	C	C
SNPO157_31	T	G	T	G	C	C
SNPO157_32	T	G	T	G	C	C
SNPO157_33	T	G	T	G	C	T
SNPO157_34	T	G	C	G	C	T
SNPO157_35	T	G	C	G	C	T
SNPO157_36	T	G	T	G	C	C
SNPO157_37	T	G	C	G	C	T
SNPO157_38	T	G	T	G	C	C
SNPO157_39	T	G	C	G	C	T

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4109.SNP-312	ECs4130.SNP-624	ECs4130.SNP-867	ECs4204.SNP-380	ECs4204.SNP-618	ECs4226.SNP-738	ECs4259.SNP-1680
493/89	T	C	G	A	A	T	T
86-24	T	C	A	A	G	C	T
87-14	T	C	A	A	G	C	T
CB9615	T	C	G	A	G	C	C
EC4115	T	C	A	A	G	C	T
EDL933	T	T	A	A	G	C	T
LSU-61	T	C	G	C	G	C	T
Sakai	T	T	A	A	G	C	T
TB182A	T	C	G	A	G	C	C
TW14359	T	C	A	A	G	C	T
TW14588	T	T	A	A	G	C	T
SNPO157_01	T	C	A	A	G	C	T
SNPO157_02	T	C	A	A	G	C	T

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4109.SNP -312	ECs4130.SNP -624	ECs4130.SNP -867	ECs4204.SNP -380	ECs4204.SNP -618	ECs4226.SNP -738	ECs4259.SNP -1680
SNPO157_03	T	C	G	A	A	T	T
SNPO157_04	T	C	A	A	G	C	T
SNPO157_05	T	C	A	A	G	C	T
SNPO157_06	T	C	A	A	G	C	T
SNPO157_07	T	C	A	A	G	C	T
SNPO157_08	T	C	A	A	G	C	T
SNPO157_09	T	C	G	A	A	T	T
SNPO157_10	T	C	A	A	G	C	T
SNPO157_11	T	C	G	A	A	T	T
SNPO157_12	T	C	G	A	A	T	T
SNPO157_13	T	C	A	A	G	C	T
SNPO157_14	T	C	A	A	G	C	T
SNPO157_15	T	C	A	A	G	C	T
SNPO157_16	T	C	A	A	G	C	T
SNPO157_17	T	C	A	A	G	C	T
SNPO157_18	T	C	A	A	G	C	T
SNPO157_19	T	C	A	A	G	C	T
SNPO157_20	T	T	A	A	G	C	T
SNPO157_21	T	C	G	A	A	T	T
SNPO157_22	T	C	A	A	G	C	T
SNPO157_23	T	T	A	A	G	C	T
SNPO157_24	T	C	A	A	G	C	T
SNPO157_25	T	C	A	A	G	C	T
SNPO157_26	T	C	A	A	G	C	T
SNPO157_27	T	C	A	A	G	C	T
SNPO157_28	T	C	A	A	G	C	T
SNPO157_29	T	C	A	A	G	C	T
SNPO157_30	T	C	A	A	G	C	T
SNPO157_31	T	C	A	A	G	C	T
SNPO157_32	T	C	A	A	G	C	T
SNPO157_33	T	C	A	A	G	C	T
SNPO157_34	T	C	G	A	A	T	T
SNPO157_35	T	C	G	A	A	T	T
SNPO157_36	T	C	G	A	G	C	T
SNPO157_37	T	C	G	A	A	T	T
SNPO157_38	T	C	A	A	G	C	T
SNPO157_39	C	C	G	A	A	T	T

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4265.SNP -609	ECs4359.SNP -1726	ECs4359.SNP -1519	ECs4364.SNP -1079	ECs4411.SNP -590	ECs4411.SNP -366	ECs4411.SNP -324
493/89	G	G	G	G	G	G	C
86-24	A	G	G	G	G	A	C
87-14	A	G	G	G	G	A	C
CB9615	G	A	G	A	G	G	C
EC4115	A	G	A	G	G	A	C
EDL933	A	G	G	G	G	A	C
LSU-61	G	G	G	G	G	G	C
Sakai	A	G	G	G	G	A	C
TB182A	G	A	G	A	G	G	C
TW14359	A	G	A	G	G	A	C
TW14588	A	G	G	G	G	A	C
SNPO157_01	A	G	G	G	G	A	C
SNPO157_02	A	G	A	G	G	A	C
SNPO157_03	G	G	G	G	G	G	C
SNPO157_04	A	G	G	G	G	A	C
SNPO157_05	A	G	G	G	G	A	C
SNPO157_06	A	G	G	G	G	A	C
SNPO157_07	A	G	A	G	G	A	C
SNPO157_08	A	G	G	G	G	A	C
SNPO157_09	G	G	G	G	G	G	C
SNPO157_10	A	G	G	G	G	A	C
SNPO157_11	G	G	G	G	G	G	C
SNPO157_12	G	G	G	G	G	G	C
SNPO157_13	A	G	A	G	G	A	C

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4265.SNP -609	ECs4359.SNP -1726	ECs4359.SNP -1519	ECs4364.SNP -1079	ECs4411.SNP -590	ECs4411.SNP -366	ECs4411.SNP -324
SNPO157_14	A	G	G	G	G	A	C
SNPO157_15	A	G	G	G	G	A	C
SNPO157_16	A	G	G	G	G	A	C
SNPO157_17	A	G	G	G	G	A	C
SNPO157_18	A	G	G	G	G	A	C
SNPO157_19	A	G	G	G	G	A	C
SNPO157_20	A	G	G	G	G	A	C
SNPO157_21	G	G	G	G	G	G	C
SNPO157_22	A	G	G	G	G	A	C
SNPO157_23	A	G	G	G	G	A	C
SNPO157_24	A	G	G	G	G	A	C
SNPO157_25	A	G	G	G	G	A	C
SNPO157_26	A	G	G	G	G	A	C
SNPO157_27	A	G	G	G	G	A	C
SNPO157_28	A	G	G	G	A	A	C
SNPO157_29	A	G	G	G	G	A	C
SNPO157_30	A	G	G	G	G	A	C
SNPO157_31	A	G	G	G	A	A	C
SNPO157_32	A	G	G	G	G	A	C
SNPO157_33	A	G	G	G	A	A	C
SNPO157_34	G	G	G	G	G	G	C
SNPO157_35	G	G	G	G	G	G	C
SNPO157_36	A	G	G	G	G	A	T
SNPO157_37	G	G	G	G	G	G	C
SNPO157_38	A	G	G	G	G	A	C
SNPO157_39	G	G	G	G	G	G	C

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4418.SNP- 467	ECs4418.SNP- 582	ECs4424.SNP- 192	ECs4454.SNP- 165	ECs4454.SNP- 402	ECs4493.SNP- 509	ECs4514.SNP- 684
493/89	G	T	T	A	C	A	C
86-24	G	G	T	G	C	A	C
87-14	G	G	T	G	C	A	A
CB9615	G	G	C	A	C	A	C
EC4115	G	G	T	G	C	A	C
EDL933	G	G	T	G	C	A	C
LSU-61	G	G	T	A	C	A	C
Sakai	G	G	T	G	C	A	C
TB182A	G	G	C	A	A	A	C
TW14359	G	G	T	G	C	A	C
TW14588	G	G	T	G	C	A	C
SNPO157_01	G	G	T	G	C	A	A
SNPO157_02	G	G	T	G	C	A	C
SNPO157_03	G	T	T	A	C	A	C
SNPO157_04	G	G	T	G	C	A	A
SNPO157_05	G	G	T	G	C	A	C
SNPO157_06	G	G	T	G	C	A	C
SNPO157_07	G	G	T	G	C	A	C
SNPO157_08	G	G	T	G	C	A	C
SNPO157_09	A	T	T	A	C	A	C
SNPO157_10	G	G	T	G	C	A	A
SNPO157_11	G	T	T	A	C	A	C
SNPO157_12	G	T	T	A	C	A	C
SNPO157_13	G	G	T	G	C	A	C
SNPO157_14	G	G	T	G	C	A	A
SNPO157_15	G	G	T	G	C	A	A
SNPO157_16	G	G	T	G	C	A	A
SNPO157_17	G	G	T	G	C	A	A
SNPO157_18	G	G	T	G	C	A	C
SNPO157_19	G	G	T	G	C	A	A
SNPO157_20	G	G	T	G	C	A	C
SNPO157_21	G	T	T	A	C	A	C
SNPO157_22	G	G	T	G	C	A	C
SNPO157_23	G	G	T	G	C	A	C
SNPO157_24	G	G	T	G	C	A	C

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4418.SNP-467	ECs4418.SNP-582	ECs4424.SNP-192	ECs4454.SNP-165	ECs4454.SNP-402	ECs4493.SNP-509	ECs4514.SNP-684
SNPO157_25	G	G	T	G	C	A	C
SNPO157_26	G	G	T	G	C	C	A
SNPO157_27	G	G	T	G	C	A	C
SNPO157_28	G	G	T	G	C	A	C
SNPO157_29	G	G	T	G	C	A	A
SNPO157_30	G	G	T	G	C	A	C
SNPO157_31	G	G	T	G	C	A	C
SNPO157_32	G	G	T	G	C	A	C
SNPO157_33	G	G	T	G	C	A	C
SNPO157_34	G	T	T	A	C	A	C
SNPO157_35	G	T	T	A	C	A	C
SNPO157_36	G	G	T	G	C	A	C
SNPO157_37	G	T	T	A	C	A	C
SNPO157_38	G	G	T	G	C	A	C
SNPO157_39	G	T	T	A	C	A	C

Strain ID	Locus tag (ORF designation) SNP position within ORF							
	ECs4601.SNP-763	ECs4689.SNP-683	ECs4711.SNP-57	ECs4841.SNP-717	ECs4861.SNP-471	ECs4872.SNP-261	ECs4920.SNP-568	
493/89	T	T	G	C	C	G	G	
86-24	C	T	G	C	C	G	G	
87-14	C	G	G	C	C	G	G	
CB9615	C	T	A	C	C	G	A	
EC4115	C	G	G	C	C	G	G	
EDL933	C	T	G	C	C	G	G	
LSU-61	C	T	G	C	C	G	G	
Sakai	C	T	G	C	C	G	G	
TB182A	C	T	A	C	C	G	A	
TW14359	C	G	G	C	C	G	G	
TW14588	C	T	G	C	C	G	G	
SNPO157_01	C	G	G	C	C	G	G	
SNPO157_02	C	G	G	C	C	G	G	
SNPO157_03	T	T	G	C	C	G	G	
SNPO157_04	C	G	G	C	C	G	G	
SNPO157_05	C	T	G	C	C	G	G	
SNPO157_06	C	T	G	C	C	G	G	
SNPO157_07	C	G	G	C	C	G	G	
SNPO157_08	C	G	G	C	C	G	G	
SNPO157_09	T	T	G	C	C	G	G	
SNPO157_10	C	G	G	C	C	G	G	
SNPO157_11	T	T	G	C	C	G	G	
SNPO157_12	T	T	G	C	C	G	G	
SNPO157_13	C	G	G	C	C	G	G	
SNPO157_14	C	G	G	C	C	G	G	
SNPO157_15	C	G	G	C	C	G	G	
SNPO157_16	C	G	G	C	C	G	G	
SNPO157_17	C	G	G	C	C	G	G	
SNPO157_18	C	T	G	C	C	G	G	
SNPO157_19	C	G	G	C	C	G	G	
SNPO157_20	C	T	G	C	C	G	G	
SNPO157_21	T	T	G	C	C	G	G	
SNPO157_22	C	T	G	C	C	G	G	
SNPO157_23	C	T	G	C	T	G	G	
SNPO157_24	C	G	G	C	T	G	G	
SNPO157_25	C	T	G	C	C	G	G	
SNPO157_26	C	G	G	C	C	G	G	
SNPO157_27	C	T	G	C	C	G	G	
SNPO157_28	C	T	G	C	C	G	G	
SNPO157_29	C	G	G	C	C	G	G	
SNPO157_30	C	T	G	C	C	G	G	
SNPO157_31	C	T	G	C	C	G	G	
SNPO157_32	C	T	G	C	C	G	G	
SNPO157_33	C	T	G	C	C	G	G	
SNPO157_34	T	T	G	C	C	G	G	
SNPO157_35	T	T	G	C	C	G	G	

Strain ID	Locus tag (ORF designation) SNP position within ORF						
	ECs4601.SNP -763	ECs4689.SNP -683	ECs4711.SNP -57	ECs4841.SNP -717	ECs4861.SNP -471	ECs4872.SNP -261	ECs4920.SNP -568
SNPO157_36	C	T	G	C	C	G	G
SNPO157_37	T	T	G	C	C	G	G
SNPO157_38	C	T	G	C	C	G	G
SNPO157_39	T	T	G	C	C	G	G

Strain ID	Locus tag (ORF designation) SNP position within ORF			
	ECs5210.SNP-308	ECs5210.SNP-366	ECs5222.SNP-344	ECs5222.SNP-70
493/89	G	T	T	C
86-24	G	T	A	C
87-14	G	T	A	T
CB9615	G	C	A	C
EC4115	G	T	A	C
EDL933	G	T	A	C
LSU-61	G	T	A	C
Sakai	A	T	A	C
TB182A	G	C	A	C
TW14359	G	T	A	C
TW14588	A	T	A	C
SNPO157_01	G	T	A	C
SNPO157_02	G	T	A	C
SNPO157_03	G	T	T	C
SNPO157_04	G	T	A	C
SNPO157_05	G	T	A	C
SNPO157_06	G	T	A	C
SNPO157_07	G	T	A	C
SNPO157_08	G	T	A	C
SNPO157_09	G	T	T	C
SNPO157_10	G	T	A	C
SNPO157_11	G	T	T	C
SNPO157_12	G	T	T	C
SNPO157_13	G	T	A	C
SNPO157_14	G	T	A	C
SNPO157_15	G	T	A	C
SNPO157_16	G	T	A	C
SNPO157_17	G	T	A	C
SNPO157_18	G	T	A	C
SNPO157_19	G	T	A	C
SNPO157_20	A	T	A	C
SNPO157_21	G	T	T	C
SNPO157_22	G	T	A	C
SNPO157_23	G	T	A	C
SNPO157_24	G	T	A	C
SNPO157_25	G	T	A	C
SNPO157_26	G	T	A	C
SNPO157_27	G	T	A	C
SNPO157_28	G	T	A	C
SNPO157_29	G	T	A	C
SNPO157_30	G	T	A	C
SNPO157_31	G	T	A	C
SNPO157_32	G	T	A	C
SNPO157_33	G	T	A	C
SNPO157_34	G	T	T	C
SNPO157_35	G	T	T	C
SNPO157_36	G	T	A	C
SNPO157_37	G	T	T	C
SNPO157_38	G	T	A	C
SNPO157_39	G	T	T	C

Technical Appendix Table 3. List of all 111 SNPs discovered in this study

ORF designation	ORF orientation within the Genome	SNP Position within the ORF	SNP Position within the Genome	Reference Nucleotide	Variant Nucleotide	Reference Codon	Variant Codon	Reference Amino acid	Variant Amino acid	Codon Effect*
ECs0076	-	1212	84260	C	T	TTG	TTA	L	L	syn
ECs0076	-	1164	84308	T	C	GCA	GCG	A	A	syn
ECs0093	+	687	103694	C	T	GCG	GGT	G	G	syn
ECs0100	+	406	111567	G	A	GTT	ATT	V	I	non-syn
ECs0204	-	573	233638	C	T	CTG	CTA	L	L	syn
ECs0406	+	494	432014	C	T	CCT	CTT	P	L	non-syn
ECs0444	+	260	472489	C	T	CCA	CTA	P	L	non-syn
ECs0502	+	342	535275	T	G	GGT	GGG	G	G	syn
ECs0523	+	1134	559120	C	T	GTC	GTT	V	V	syn
ECs0552	-	186	609385	C	T	ATG	ATA	M	I	non-syn
ECs0614	+	138	684625	C	T	GGC	GGT	G	G	syn
ECs0614	+	283	684770	T	A	TGG	AGG	W	R	non-syn
ECs0647	+	216	723657	T	C	GAT	GAC	D	D	syn
ECs0647	+	306	723747	C	T	GCC	GCT	A	A	syn
ECs0647	+	556	723997	C	G	CAA	GAA	Q	E	non-syn
ECs0905	-	453	990246	C	T	CCG	CCA	P	P	syn
ECs0905	-	217	990482	G	C	CCG	GCG	P	A	non-syn
ECs0917	+	578	1004607	C	T	CCG	CTG	P	L	non-syn
ECs0955	-	745	1038649	C	A	GTG	TTG	V	L	non-syn
ECs0955	-	719	1038675	C	T	CGC	CAC	R	H	non-syn
ECs0998	+	375	1098382	C	T	CCC	CCT	P	P	syn
ECs1044	-	2080	1151284	A	G	TTA	CTA	L	L	syn
ECs1044	-	1773	1151591	C	T	GTG	GTA	V	V	syn
ECs1046	+	319	1154260	C	T	CTG	TTG	L	L	syn
ECs1046	+	358	1154299	C	A	CTG	ATG	L	M	non-syn
ECs1137	-	524	1222818	A	G	ATG	ACG	M	T	non-syn
ECs1137	-	230	1223112	C	T	AGT	AAT	S	N	non-syn
ECs1148	-	1014	1233518	C	A	ATG	ATT	M	I	non-syn
ECs1148	-	940	1233592	G	A	CGT	TGT	R	C	non-syn
ECs1729	+	786	1725872	C	T	ACC	ACT	T	T	syn
ECs1741	-	2155	1740010	G	A	CTG	TTG	L	L	syn
ECs1833	-	474	1818962	C	T	CCG	CCA	P	P	syn
ECs1871	-	1109	1863935	A	G	TTG	TCG	L	S	non-syn
ECs1871	-	978	1864066	G	A	CGC	CGT	R	R	syn
ECs1878	+	829	1872868	G	T	GGC	TGC	G	C	non-syn
ECs2022	-	793	2007430	T	A	ACC	TCC	T	S	non-syn
ECs2045	+	677	2029256	C	T	CCG	CTG	P	L	non-syn
ECs2087	-	253	2079029	T	C	ATG	GTG	M	V	non-syn
ECs2320	+	486	2290138	A	G	CTA	CTG	L	L	syn
ECs2320	+	719	2290371	A	T	GAA	GTA	E	V	non-syn
ECs2435	+	123	2410770	G	A	GTG	GTA	V	V	syn
ECs2435	+	458	2411105	T	A	ATC	AAC	I	N	non-syn
ECs2454	-	1064	2430431	G	A	TCT	TTT	S	F	non-syn
ECs2454	-	693	2430802	A	G	CGT	CGC	R	R	syn
ECs2582	-	2038	2556900	G	A	CGT	TGT	R	C	non-syn
ECs2583	-	73	2559990	T	G	ATG	CTG	M	L	non-syn
ECs2589	-	804	2566177	A	C	GTT	GTG	V	V	syn
ECs2589	-	771	2566210	A	C	GCT	GCG	A	A	syn
ECs3043	-	721	2975941	C	T	GTA	ATA	V	I	non-syn
ECs3059	-	1440	2994900	A	C	CCT	CCG	P	P	syn
ECs3076	+	311	3015872	T	C	CTT	CCT	L	P	non-syn
ECs3076	+	573	3016134	T	G	ATT	ATG	I	M	non-syn
ECs3076	+	1413	3016974	G	A	GCG	GCA	A	A	syn
ECs3076	+	1689	3017250	G	A	CAG	CAA	Q	Q	syn
ECs3207	-	291	3167678	C	A	CCG	CCT	P	P	syn
ECs3225	-	1098	3185787	C	T	GCG	GCA	A	A	syn
ECs3225	-	904	3185981	G	A	CGC	TGC	R	C	non-syn
ECs3231	+	1105	3194248	C	T	CGC	TGC	R	C	non-syn
ECs3246	-	1113	3209201	C	T	CCG	CCA	P	P	syn
ECs3246	-	743	3209571	A	G	TTT	TCT	F	S	non-syn
ECs3300	+	458	3270415	G	A	GGC	GAC	G	D	non-syn
ECs3300	+	855	3270812	T	A	GGT	GGA	G	G	syn
ECs3377	-	522	3357424	A	C	GAT	GAG	D	E	non-syn
ECs3377	-	226	3357720	T	G	AAA	CAA	K	Q	non-syn
ECs3377	-	127	3357819	C	T	GCA	ACA	A	T	non-syn

ORF designation	ORF orientation within the Genome	SNP Position within the ORF	SNP Position within the Genome	Reference Nucleotide	Variant Nucleotide	Reference Codon	Variant Codon	Reference Amino acid	Variant Amino acid	Codon Effect*
ECs3464	-	297	3460973	G	T	CCC	CCA	P	P	syn
ECs3464	-	227	3461043	C	A	CGA	CTA	R	L	non-syn
ECs3564	+	786	3554521	C	A	ACC	ACA	T	T	syn
ECs3676	-	147	3674715	C	T	GGG	GGA	G	G	syn
ECs3746	+	984	3749342	C	T	GCG	GGT	G	G	syn
ECs3746	+	1136	3749494	A	C	CAA	CCA	Q	P	non-syn
ECs3773	+	1100	3782791	A	C	GAG	GCG	E	A	non-syn
ECs3810	-	429	3821818	G	A	GAC	GAT	D	D	syn
ECs3833	-	365	3841958	T	C	GAC	GGC	D	G	non-syn
ECs3839	+	793	3846816	G	A	GCC	ACC	A	T	non-syn
ECs3994	-	1663	4000385	T	C	AAA	GAA	K	E	non-syn
ECs4047	-	272	4055003	G	A	GCC	GTC	A	V	non-syn
ECs4109	-	873	4118633	C	A	GCG	GCT	A	A	syn
ECs4109	-	812	4118694	C	T	GGT	GAT	G	D	non-syn
ECs4109	-	312	4119194	T	C	CAA	CAG	Q	Q	syn
ECs4130	+	624	4143190	T	C	GGT	GGC	G	G	syn
ECs4130	+	867	4143433	A	G	CTA	CTG	L	L	syn
ECs4204	+	380	4203487	A	C	GAA	GCA	E	A	non-syn
ECs4204	+	618	4203725	G	A	CTG	CTA	L	L	syn
ECs4226	-	738	4225097	C	T	CTG	CTA	L	L	syn
ECs4259	-	1680	4262902	T	C	CCA	CCG	P	P	syn
ECs4265	+	609	4271823	A	G	GTA	GTG	V	V	syn
ECs4359	-	1726	4362239	G	A	CGC	TGC	R	C	non-syn
ECs4359	-	1519	4362446	G	A	CCT	TCT	P	S	non-syn
ECs4364	-	1079	4370817	G	A	CCT	CTT	P	L	non-syn
ECs4411	-	590	4431800	G	A	CCG	CTG	P	L	non-syn
ECs4411	-	366	4432024	A	G	GCT	GCC	A	A	syn
ECs4411	-	324	4432066	C	T	GTG	GTA	V	V	syn
ECs4418	+	467	4440826	G	A	GGA	GAA	G	E	non-syn
ECs4418	+	582	4440941	G	T	ATG	ATT	M	I	non-syn
ECs4424	-	192	4449503	T	C	CTA	CTG	L	L	syn
ECs4454	+	165	4483743	G	A	CAG	CAA	Q	Q	syn
ECs4454	+	402	4483980	C	A	AGC	AGA	S	R	non-syn
ECs4493	-	509	4536406	A	C	GTT	GGT	V	G	non-syn
ECs4514	+	684	4557273	C	A	GCG	GGA	G	G	syn
ECs4601	-	763	4630237	C	T	GTC	AGC	G	S	non-syn
ECs4689	+	683	4725442	T	G	GTC	GGC	V	G	non-syn
ECs4711	+	57	4754139	G	A	CTG	CTA	L	L	syn
ECs4841	+	717	4908228	C	T	ACC	ACT	T	T	syn
ECs4861	-	471	4925478	C	T	CCG	CCA	P	P	syn
ECs4872	+	261	4943251	G	A	GCG	GCA	A	A	syn
ECs4920	+	568	5007108	G	A	GCT	ACT	A	T	non-syn
ECs5210	+	308	5311572	A	G	CAC	CGC	H	R	non-syn
ECs5210	+	366	5311630	T	C	ACT	ACC	T	T	syn
ECs5222	-	344	5327590	A	T	CTG	CAG	L	Q	non-syn
ECs5222	-	70	5327864	C	T	GTG	ATG	V	M	non-syn

*syn, synonymous; non-syn, non-synonymous